



UNIVERSITI PUTRA MALAYSIA

***MOLECULAR EPIDEMIOLOGY AND GENETIC DIVERSITY OF  
PLASMODIUM KNOWLESI INFECTING LONG-TAILED MACAQUES  
(MACACA FASCICULARIS RAFFLES) ON THE WEST COAST OF  
PENINSULAR MALAYSIA***

LEE COL LIN

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By

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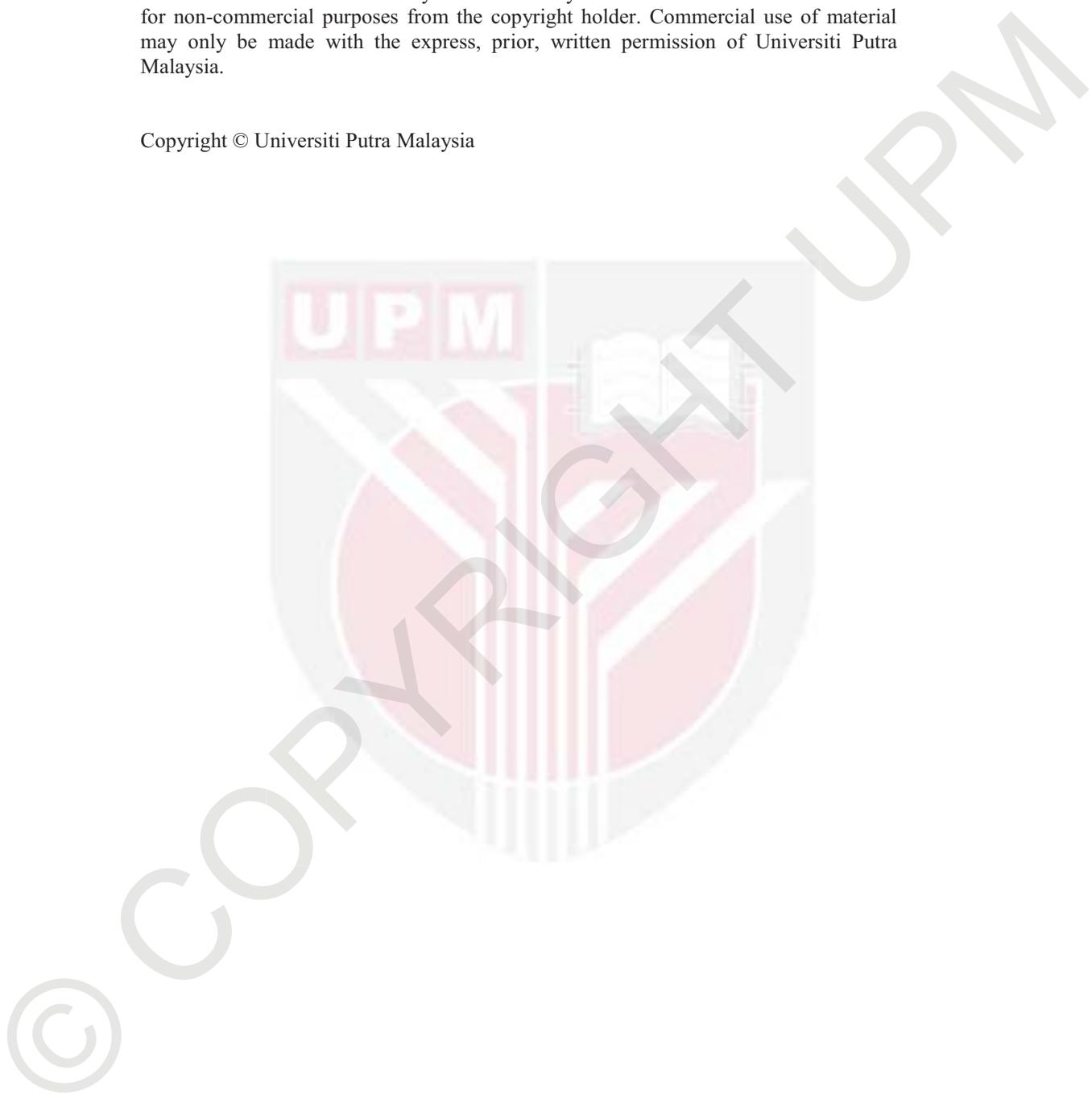


**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Master of Science**

**December 2015**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of  
the requirements for the degree of Master of Science

**MOLECULAR EPIDEMIOLOGY AND GENETIC DIVERSITY OF  
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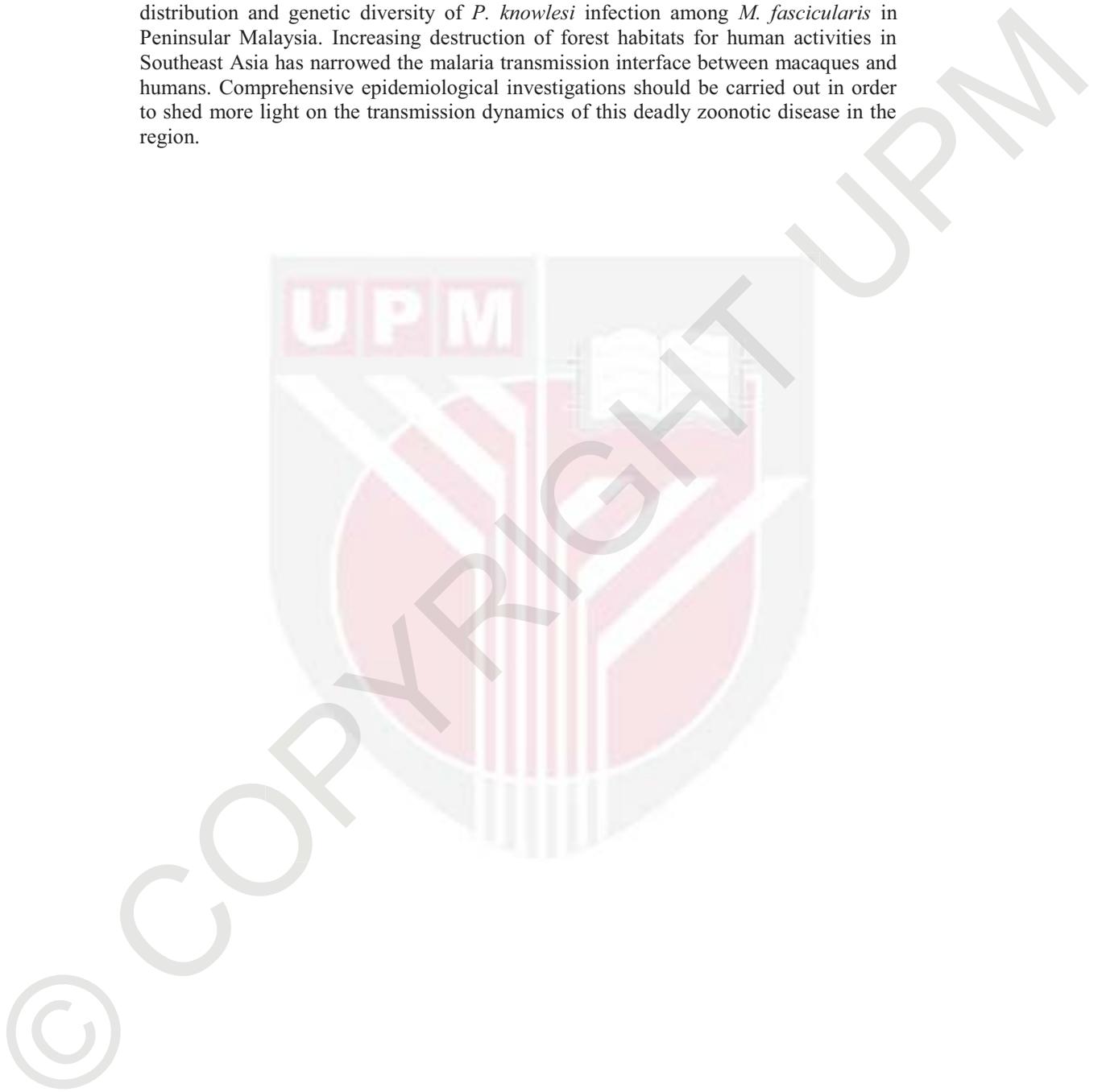
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**December 2015**

**Chairman: Reuben Sunil Kumar Sharma, PhD  
Faculty: Veterinary Medicine**

This study was undertaken to ascertain the molecular epidemiology of *Plasmodium knowlesi* infecting wild Long-tailed macaques (*Macaca fascicularis*) on the west coast of Peninsular Malaysia, and to determine its spatial distribution and risk factors for infection. Genetic analysis based on the Circumsporozoite Protein (CSP) gene of *P. knowlesi* was done to infer the genetic assemblage of the parasite. A total of 781 blood samples were collected from wild *M. fascicularis* captured from 77 locations representing four different habitats; urban, sub-urban, plantation/orchards and secondary forest. Five states on the west coast of the country were sampled, namely, Penang, Perak, Selangor, Negeri Sembilan and Melaka, and the area was arbitrary divided into three zones. Ten human infected blood samples from Kuala Kubu Baru, Selangor were included in the study. Screening for *P. knowlesi* infection was conducted using nested PCR targeting the 18S SSU rRNA and phylogenetic characterization was done using the CSP gene. Prevalence of infection was 13.4% (15.1% male and 11.1% female) with Zone 1 exhibiting the highest prevalence (29.8%), followed by Zone 3 (15.6%) and Zone 2 (5.3%). Macaques inhabiting the plantation/orchards were the most infected (18.2%), followed by sub-urban (16.5%), secondary forest (15.7%), and urban (9.8%) areas. The adult macaques showed the highest prevalence of *P. knowlesi* infection (16.6%), followed by the juveniles (13.1%) and sub-adults (9.7%). Spatial distribution analysis revealed two hotspots of infection in the country; on the northwest (Penang and north Perak) and southwest (Negeri Sembilan). Risk factor analysis showed that all putative factors except gender, posed a risk for infection with this parasite among the macaques. Stepwise binary logistic regression analysis revealed that macaques from Zone 1, sub-urban and plantation/orchards, and adults, have higher risk of infections compared to conspecifics in the other categories. Genotypic analyses of 192 *P. knowlesi* CSP gene sequences (178 from macaques and 14 from humans) produced 25 different haplotypes with 14 polymorphic sites. The overall nucleotide diversity ( $\pi = 0.0196$ ) and haplotype diversity ( $H_d = 0.836$ ) was high, but genetic differentiation between the zones and habitats was low ( $F_{ST} < 0.05$ ). The three most commonly encountered haplotypes PkMH18 ( $f = 62$ ), PkMH13 ( $f = 26$ ) and PkMH04 ( $f = 18$ ), were widely distributed across the sampling locations, with the former predominating in the northwest region of the country. All eight haplotypes from

humans were found to be identical with that obtained from the macaques. Phylogenetic analysis clustered macaque and human isolates from within the Southeast Asia region. This study represents the first attempt to elucidate the molecular epidemiology, spatial distribution and genetic diversity of *P. knowlesi* infection among *M. fascicularis* in Peninsular Malaysia. Increasing destruction of forest habitats for human activities in Southeast Asia has narrowed the malaria transmission interface between macaques and humans. Comprehensive epidemiological investigations should be carried out in order to shed more light on the transmission dynamics of this deadly zoonotic disease in the region.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk ijazah Master Sains

**EPIDEMIOLOGI MOLEKUL DAN KEPELBAGAIAN GENETIK  
*PLASMODIUM KNOWLESI* YANG MENJANGKITI KERA (*MACACA FASCICULARIS*) DI PANTAI BARAT SEMENANJUNG MALAYSIA**

Oleh

**LEE COL LIN**

**Disember 2015**

**Pengerusi: Reuben Sunil Kumar Sharma, PhD**

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Kajian ini telah dijalankan untuk menentukan epidemiologi molekula jangkitan *Plasmodium knowlesi* dalam kera liar (*Macaca fascicularis*) di kawasan pantai barat Semenanjung Malaysia dan untuk menentukan taburan spatial dan faktor risiko yang berkaitan dengan jangkitan. Analisis genetik berdasarkan gen Circumsporozoite Protein (CSP) *P. knowlesi* dilakukan untuk membuat kesimpulan himpunan genetik parasit. Sebanyak 781 sampel darah telah dikumpulkan dari kera liar, *M. fascicularis* yang ditangkap dari 77 lokasi yang mewakili empat habitat yang berbeza; bandar, pinggir bandar, ladang/kebun dan hutan sekunder. Lima negeri di pantai barat negara ini telah disampel, iaitu, Pulau Pinang, Perak, Selangor, Negeri Sembilan dan Melaka, dan kawasan itu dibahagi kepada tiga zon. Sepuluh sampel darah manusia dari Kuala Kubu Baru, Selangor yang sah dijangkiti *P. knowlesi* telah disertakan dalam kajian ini. Saringan untuk jangkitan *P. knowlesi* telah dijalankan dengan menggunakan kaedah PCR tersarang yang mensasarkan 18S SSU rRNA dan pencirian filogenetik telah dilakukan dengan menggunakan gen CSP. Prevalen jangkitan *P. knowlesi* adalah 13.4% (15.1% jantan dan 11.1% betina) dengan Zon 1 menunjukkan prevalen tertinggi (29.8%), diikuti oleh Zon 3 (15.6%) dan Zon 2 (5.3%). Jangkitan dalam kera yang datang dari ladang/kebun adalah paling tinggi (18.2%), diikuti oleh kawasan sub-bandar (16.5%), hutan sekunder (15.7%), dan bandar (9.8%). Kera dewasa menunjukkan kelaziman jangkitan *P. knowlesi* tertinggi (16.6%), diikuti oleh remaja (13.1%) dan sub-dewasa (9.7%). Analisis taburan spatial mendedahkan dua titik panas jangkitan di negara ini; di barat laut (Pulau Pinang dan utara Perak) dan barat daya (Negeri Sembilan). Analisis faktor risiko menunjukkan bahawa semua faktor, kecuali jantina, adalah risiko jangkitan parasit di antara kera. Analisis binari regresi logistik langkah demi langkah mendedahkan bahawa kera dari Zon 1, sub-bandar dan ladang/kebun, dan dewasa, mempunyai risiko yang lebih tinggi berbanding dengan kera dalam kategori lain. Analisis genotip berdasarkan 192 jujukan gen CSP *P. knowlesi* (178 dari kera dan 14 dari manusia) menghasilkan 25 haplotip berbeza dengan 14 tapak polimorfik. Kepelbagaiannya keseluruhan nukleotida ( $\pi = 0.0196$ ) dan kepelbagaiannya haplotip ( $Hd = 0.836$ ) adalah tinggi, tetapi perbezaan genetik antara zon dan habitat adalah rendah ( $F_{ST} < 0.05$ ). Tiga haplotip yang paling biasa adalah PkMH18 ( $f = 62$ ), PkMH13 ( $f = 26$ ) dan PkMH04 ( $f = 18$ ), dan mereka ditemui secara meluas di seluruh

lokasi persampelan, dengan PkMH13 mendominasi rantaui barat laut negara ini. Kesemua lapan haplotip dari sampel manusia didapati serupa dengan yang diperoleh dari kera. Analisis filogenetik mendapati bahawa isolat kera dan manusia adalah berkelompok dalam rantaui Asia Tenggara. Kajian ini merupakan percubaan pertama untuk menjelaskan epidemiologi molekul, taburan spatial dan kepelbagaian genetik *P. knowlesi* antara *M. fascicularis* di Semenanjung Malaysia. Peningkatan kemusnahan habitat hutan untuk aktiviti manusia di Asia Tenggara telah mengurangkan jurang aliran malaria antara kera dan manusia. Penyiasatan epidemiologi secara komprehensif perlu dijalankan untuk menjelaskan lebih lanjut tentang dinamik aliran penyakit zoonotik yang membawa maut di rantaui ini.



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I certify that a Thesis Examination Committee has met on 10 December 2015 to conduct the final examination of Lee Col Lin on her thesis entitled "Molecular Epidemiology and Genetic Diversity of *Plasmodium knowlesi* Infecting Long-Tailed Macaques (*Macaca fascicularis* Raffles) on the West Coast of Peninsular Malaysia" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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## LIST OF ABBREVIATIONS

°C	degree celcius
µl	microlitre
µM	micromolar
1x	one time
An	<i>Anopheles</i>
bp	base pair
C	Cystine
CaCl <sub>2</sub>	calcium chloride
cm	centimetre
COI	cytochrome oxidase subunit I
COII	cytochrome oxidase subunit II
CSP	Circumsporozoite Protein
cyt	cytochrome
Cytb	cytochrome b
D	Aspartic acid
DNA	deoxyribonucleic acid
dNTP	deoxynucleotide triphosphate
E	Glutamic acid
EDTA	ethylenediamine tetraacetic acid
ExPasy	Expert Protein Analysis System
F	Phenylalanine
F <sub>D</sub>	Fu and Li's D
F <sub>ST</sub>	genetic differentiation
ft	feet
G	Glycine
GAG	glycosaminoglycan
GPI	glycosylphosphatidylinositol
GPS	Global Positioning System
h	hour
Hd	haplotype diversity
HRP-II	histidine-rich protein II
HSPG	heparin sulphate proteoglycan
I	Isoleucine
K	Lysine
kb	kilobase
kDa	kilo Dalton
kg	kilogram
km	kilometer
L	Leucine
LAMP	loop-mediated isothermal amplification
LB	Lysogeny Broth
lb	pound
LDH	lactate dehydrogenase
M	Methionine
m	metre
mg	milligram
MgCL <sub>2</sub>	magnesium chloride
min	minute

ML	Maximum Likelihood
ml	millilitre
mM	millimolar
mm	millimetre
MOH	Ministry of Health
MSP	merozoite surface protein
MSP-1	merozoite surface protein-1
mtDNA	mitochondria deoxyribonucleic acid
N	Asparagine
NCBI	National Center for Biotechnology Information
ng	nanogram
NJ	Neighbour-joining
nm	nanometer
OR	odds ratio
ORF	open reading frame
P	Proline
<i>P.</i>	<i>Plasmodium</i>
PCR	polymerase chain reaction
PERHILITAN	Department of Wildlife and National Parks Malaysia
pi	nucleotide diversity
qPCR	quantitative polymerase chain reaction
RDT	rapid diagnostic test
RNase	ribonuclease
rpm	revolution per minute
S	Serine
sec	second
SSU rRNA	small subunit ribosomal RNA
TAE	Tris-acetic acid-EDTA
$T_D$	Tajima's D
U	unit
UV	ultraviolet
V	Alanine
V	volt
w/v	weight over volume
WHO	World Health Organization

## CHAPTER 1

### GENERAL INTRODUCTION

Malaria is a disease of public health importance and is caused by the blood protozoan parasites of the genus *Plasmodium* which is transmitted by *Anopheles* mosquitoes. The parasite is known to infect humans and a wide range of animal taxa (Coatney *et al.*, 1971; Rich and Ayala, 2006; Ramasamy, 2014). Human malaria is widely distributed across tropical and sub-tropical regions, covering 97 countries, including Asia, Africa and South America (World Health Organization, 2014). Latest statistical reports by the World Health Organization (2014) indicate that 1.2 billion people are living in areas with high risk of malaria transmission, accounting for over 584,000 deaths of which 78% involve children aged under 5 years (WHO, 2014).

For over 100 years since the discovery of the *Plasmodium* parasites, numerous studies have been carried out and five species are known to infect humans; *P. falciparum*, *P. vivax*, *P. malariae*, *P. ovale* and *P. knowlesi* (Eyles *et al.*, 1960; Chin *et al.*, 1965; Singh *et al.*, 2004; Ramasamy, 2014; Ta *et al.*, 2014; White *et al.*, 2014). Among these, *P. vivax* is reported to be the most widely distributed species of human malaria especially in Asia and South America, whereas *P. falciparum* predominates only in sub-Saharan Africa region but is the most lethal cause of infections (Eyles *et al.*, 1960; Chin *et al.*, 1965; Guerra *et al.*, 2008; 2010; Hay *et al.*, 2010; Tilley *et al.*, 2011).

Over 30 *Plasmodium* species have been described in non-human primate thus far, infecting a wide range of monkeys, apes and lemurs (Kantele and Jokiranta, 2011; Ramasamy, 2014). Macaques are found to harbour the most variety of non-human *Plasmodium* species, namely, *P. knowlesi*, *P. inui*, *P. cynomolgi*, *P. coatneyi* and *P. fieldi* (Coatney *et al.*, 1971; Fooden, 1982; 1994; Lee *et al.*, 2011). In Malaysia, two most common macaque species, the Long-tailed macaque (*Macaca fascicularis*) and the Pig-tailed macaque (*Macaca nemestrina*) have been commonly studied for malaria infections and are known to harbour zoonotic *Plasmodium* species (Eyles *et al.*, 1962; Coatney *et al.*, 1971; Southwick and Cadigan, 1972; Fooden, 1982; 1994; Vythilingam *et al.*, 2008; Lee *et al.*, 2011). *Macaca fascicularis* is the most abundantly distributed macaque in forested area of Southeast Asia, especially in Peninsular Malaysia (Southwick and Cadigan, 1972; Fooden, 1982; 1995). Of all known non-human primate species, *P. knowlesi* (Chin *et al.*, 1968; Antinori *et al.*, 2013), *P. inui* (Coatney, 1966) and *P. cynomolgi* (Coatney *et al.*, 1961; Eyles 1960; Schmidt *et al.*, 1961; Ta *et al.*, 2014) are proven to be zoonotic and transmissible to human via the bite of the *Anopheles* s group of mosquito.

The discovery of a large focus of *P. knowlesi* human malaria cases in the Kapit Division of Sarawak (Singh *et al.*, 2004), has evoked interest among many researchers studying the infection in humans. The polymerase chain reaction (PCR) assay using molecular markers like the small subunit ribosomal RNA (SSU rRNA) and circumsporozoite protein (CSP) gene (Singh *et al.*, 2004) is now the choice technique for the detection of malaria in primates and humans due to its sensitivity and specificity

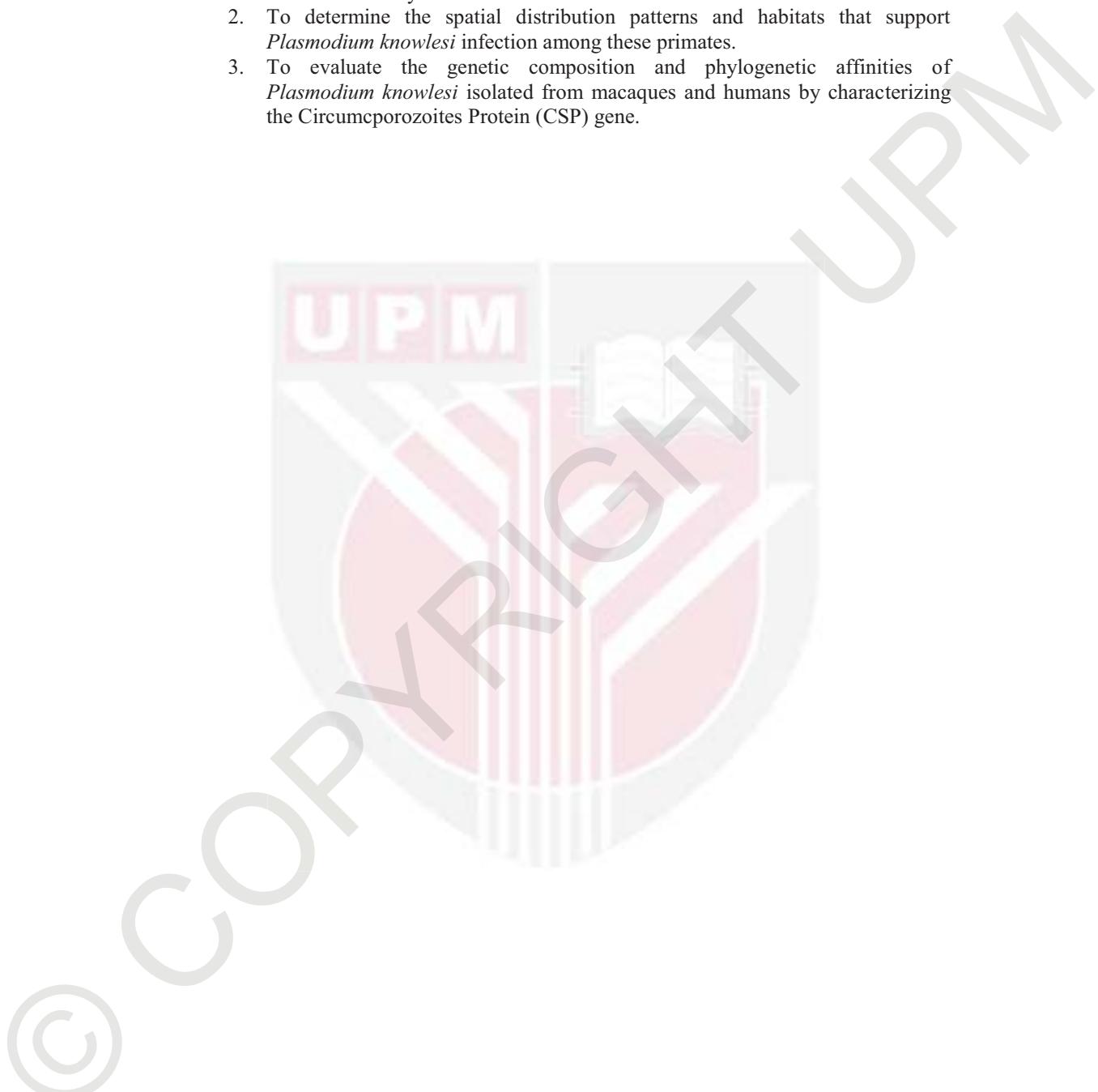
compared to traditional microscopy. Molecular tools are able to differentiate morphologically similar species like *P. malariae* and *P. knowlesi*, leading to the discovery that *P. knowlesi* is a prominent form of human malaria in many parts of Southeast Asia (Jongwutiwes *et al.*, 2004; Singh *et al.*, 2004; Cox-Singh *et al.*, 2008; Ng *et al.*, 2008; Luchavez *et al.*, 2008; Ramasamy, 2013; William *et al.*, 2013, 2014; Yusof *et al.*, 2014). A recent survey (Yusuf *et al.*, 2014) highlighted the widespread nature of human *knowlesi* infections in most states of Peninsular Malaysia, corroborating the recent statistical report by the Ministry of Health, Malaysia (2013) which demonstrated that *P. knowlesi* accounted for 38.4% of the total human malaria cases reported in 2012 in Malaysia. In Sabah, East Malaysia, a similar increase of *P. knowlesi* cases have been observed since 2014, inversely correlated with the decrease in *P. vivax* and *P. falciparum* infection in humans (William *et al.*, 2013; 2014).

The devastating effects of the disease are of major concern in Asia. As such, Malaysia together with its Asian partners has committed to strive towards a National Malaria Elimination goal by the end of 2020 (Roll Back Malaria Partnership, 2012; Sanders *et al.*, 2014). Latest statistics revealed that in Malaysia, there were 5306 malaria cases in 2011, 4725 cases in 2012, and 3810 cases in 2013; the highest among the vector borne parasitosis in the country (Ministry of Health Malaysia, 2012; WHO World Malaria Report, 2013; 2014). Apart from the traditional species that cause malaria in humans, *P. knowlesi* is now recognised as a parasite that can cause severe disease with high fatalities (Cox-Singh *et al.*, 2008; Mohamed and Roshan, 2009; William *et al.*, 2011; Barber *et al.*, 2013). It is also considered the most rampant (Daneshvar *et al.*, 2009; Singh and Daneshvar, 2010; Ministry of Health Malaysia, 2012; William *et al.*, 2013; Yusof *et al.*, 2014) and most severe (Barber *et al.*, 2013) form of human malaria in Malaysia, being responsible for over half of the malaria related deaths in the country (Rajahram *et al.*, 2012).

Over the years, several entomological studies have been conducted in Malaysia to elucidate the mosquito vectors involved in the transmission of *P. knowlesi* in human and macaques. These studies have incriminated the *Leucosphyrus* Group of mosquitoes as major vectors, including *An. latens* in Kapit, Sarawak (Vythilingam *et al.*, 2006), *An. cracens* in Pahang (Vythilingam *et al.*, 2008), *An. introlatus* in Hulu Selangor (Vythilingam *et al.*, 2014) and previously, *An. hackeri* in mangrove areas of Selangor (Wharton and Eyles, 1961). With the changing landscape in the country and the conversion of forested habitats for human-related activities and industry, it is likely that the spatial distribution patterns of both macaques and the mosquito vectors will be affected, leading to alterations in the transmission dynamics of the disease.

Previous studies on non-human primate malarias in the country have focused mainly on *Plasmodium* detection, prevalence and vector diversity, leaving paucity in areas of molecular epidemiology, population genetics and phylogenetic affinities of the parasite. The present investigation was therefore designed to contribute to the understanding of non-human primate malaria in the country, caused by the zoonotic *P. knowlesi* with the following specific objectives:

1. To ascertain the molecular epidemiology of *Plasmodium knowlesi* infecting wild Long-tailed Macaques (*Macaca fascicularis*) on the West Coast of Peninsular Malaysia.
2. To determine the spatial distribution patterns and habitats that support *Plasmodium knowlesi* infection among these primates.
3. To evaluate the genetic composition and phylogenetic affinities of *Plasmodium knowlesi* isolated from macaques and humans by characterizing the Circumsporozoites Protein (CSP) gene.



## REFERENCES

- Adams, J. C., and Tucker, R. P. (2000). The thrombospondin type 1 repeat (TSR) superfamily: Diverse proteins with related roles in neuronal development. *Developmental Dynamics*, 218(2), 280-299. doi: 10.1002/(SICI)1097-0177(200006)218:2<280::AID-DVDY4>3.0.CO;2-0
- Aldrich, C., Magini, A., Emiliani, C., Dottorini, T., Bistoni, F., Crisanti, A., and Spaccapelo, R. (2012). Roles of the amino terminal region and repeat region of the *Plasmodium berghei* circumsporozoite protein in parasite infectivity. *PloS One*, 7(2), e32524. doi:10.1371/journal.pone.0032524
- Amir, A., Sum, J. S., Lau, Y. L., Vytilingam, I., and Fong, M. Y. (2013). Colonization of *Anopheles cracens*: A malaria vector of emerging importance. *Parasit Vectors*, 6, 81. doi:10.1186/1756-3305-6-81
- Anderios, F., Mohamed, Z., Ratnam, S., Ibrahim, M. Y., and Awang, T. A. M. (2008). Detection of malaria parasites in Sabah by nested polymerase chain reaction: a focus of naturally acquired *Plasmodium knowlesi* infections. *Sains Malaysiana*, 37(2), 137-141.
- Anderios, F., Noorain, A., and Vytilingam, I. (2010). *In vivo* study of human *Plasmodium knowlesi* in *Macaca fascicularis*. *Experimental parasitology*, 124(2), 181-189. doi:10.1016/j.exppara.2009.09.009
- Anderson, T. J., Haubold, B., Williams, J. T., Estrada-Franco, J. G., Richardson, L., Mollinedo, R., Bockarie, M., Mokili, J., Mharakurwa, S., French, N., Whitworth, J., Velez, I. D., Brockman, A. H., Nosten, F., Ferreira, M. U., and Day, K. P. (2000). Microsatellite markers reveal a spectrum of population structures in the malaria parasite *Plasmodium falciparum*. *Molecular Biology and Evolution*, 17(10), 1467-1482.
- Antinori, S., Galimberti, L., Milazzo, L., and Corbellino, M. (2013). *Plasmodium knowlesi*: The emerging zoonotic parasite. *Acta Tropica*, 125(2):191-201. doi: 10.1016/j.actatropica.2012.10.008.
- Archibald, H. M. (1956). The influence of malarial infection of the placenta on the incidence of prematurity. *Bulletin of the World Health Organization*, 15(3-5), 842.
- Arez, A. P., Lopes, D., Pinto, J., Franco, A. S., Snounou, G., and Do Rosário, V. E. (2000). *Plasmodium* sp.: Optimal protocols for PCR detection of low parasite numbers from mosquito (*Anopheles* sp.) samples. *Experimental Parasitology*, 94(4), 269-272. doi:10.1006/expr.2000.4496
- Ariey, F. (2009). Towards high- throughput molecular detection of *Plasmodium*: New approaches and molecular markers. *Malaria Journal*, 12:86. doi: 10.1186/1475-2875-8-86.

- Arnott, A., Barry, A. E., and Reeder, J. C. (2012). Understanding the population genetics of *Plasmodium vivax* is essential for malaria control and elimination. *Malaria Journal*, 11(14), 10-1186.
- Azidah, A. K., Faizal, M., Lili, H. Y., and Zeehaida, M. (2014). Severe *Plasmodium knowlesi* infection with multiorgan involvement in north east peninsular Malaysia. *Tropical Biomedicine*, 31(1), 31-35.
- Baird, J. K. (2009). Malaria zoonoses. *Travel Medicine and Infectious Disease*, 7(5), 269-277.
- Bandelt, H. J., Forster, P., and Röhl, A. (1999). Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution*, 16(1), 37-48.
- Barber, B. E., William, T., Jikal, M., Jilip, J., Dhararaj, P., Menon, J., Yeo, T. W., and Anstey, N. M. (2011). *Plasmodium knowlesi* malaria in children. *Emerging Infectious Diseases*, 17(5), 814-820. doi: 10.3201/eid1705.101489
- Barber, B. E., William, T., Dhararaj, P., Anderios, F., Grigg, M. J., Yeo, T. W., and Anstey, N. M. (2012). Epidemiology of *Plasmodium knowlesi* malaria in north-east Sabah, Malaysia: family clusters and wide age distribution. *Malaria Journal*, 11(401), 10-1186. doi:10.1186/1475-2875-11-401
- Barber, B. E., William, T., Grigg, M. J., Yeo, T. W., and Anstey, N. M. (2013). Limitations of microscopy to differentiate *Plasmodium* species in a region co-endemic for *Plasmodium falciparum*, *Plasmodium vivax* and *Plasmodium knowlesi*. *Malar J*, 12(8), 10-1186. doi:10.1186/1475-2875-12-8
- Barber, B. E., William, T., Grigg, M. J., Menon, J., Auburn, S., Marfurt, J., Anstey, N. M., and Yeo, T. W. (2013). A prospective comparative study of *knowlesi*, *falciparum*, and *vivax* malaria in Sabah, Malaysia: high proportion with severe disease from *Plasmodium knowlesi* and *Plasmodium vivax* but no mortality with early referral and artesunate therapy. *Clinical Infectious Diseases*, 56(3), 383-397.doi:10.1093/cid/cis902
- Basommi, L. P. (2011). *Spatial Analysis of Malaria Epidemiology in the Amanse West District* (Doctoral dissertation, Kwame Nkrumah University of Science and Technology).
- Bayoh, M. N., and Lindsay, S. W. (2003). Effect of temperature on the development of the aquatic stages of *Anopheles gambiae* sensu stricto (Diptera: Culicidae). *Bulletin of Entomological Research*, 93: 375. doi: 10.1079/ber2003259.
- Bayoh, M. N., and Lindsay, S. W. (2004). Temperature-related duration of aquatic stages of the Afrotropical malaria vector mosquito *Anopheles gambiae* in the laboratory. *Medical and Veterinary Entomology*, 18: 174–179. doi: 10.1111/j.0269-283x.2004.00495.x
- Berry, A., Benoit-Vical, F., Fabre, R., Cassaing, S., and Magnaval, J. F. (2008). PCR-based methods to the diagnosis of imported malaria. *Parasite*, 15(3), 484-488.

- Berry, A., Iriart, X., Wilhelm, N., Valentin, A., Cassaing, S., Witkowski, B., Benoit-Vical, F., Menard, S., Olagnier, D., Fillaux, J., and Sire, S. (2011). Imported *Plasmodium knowlesi* malaria in a French tourist returning from Thailand. *The American Journal of Tropical Medicine and Hygiene*, 84(4), 535-538. doi: 10.4269/ajtmh.2011.10-0622.
- Birkenmeyer, L., Muerhoff, A. S., Dawson, G. J., and Desai, S. M. (2010). Isolation and characterization of the MSP1 genes from *Plasmodium malariae* and *Plasmodium ovale*. *The American Journal of Tropical Medicine and Hygiene*, 82(6), 996-1003. doi:10.4269/ajtmh.2010.09-0022
- Bowen, W. H., and Koch, G. (1970). Determination of age in monkeys (*Macaca irus*) on the basis of dental. *Laboratory Animals*, 4(1) 113-123. doi: 10.1258/002367770781036481
- Braima, K. A., Sum, J. S., Ghazali, A. R. M., Muslimin, M., Jeffery, J., Lee, W. C., Shaker, M. R., Elamin, A. E. M., Jamaiah, I., Lau, Y. L., and Rohela, M. (2013). Is there a risk of suburban transmission of malaria in Selangor, Malaysia?. *PloS one*, 8(10), e77924. doi: 10.1371/journal.pone.0077924
- Bronner, U., Divis, P. C., Färnert, A., and Singh, B. (2009). Swedish traveller with *Plasmodium knowlesi* malaria after visiting Malaysian Borneo. *Malaria Journal*. 8:15. doi: 10.1186/1475-2875-8-15.
- Brito, C. F. A. D., and Ferreira, M. U. (2011). Molecular markers and genetic diversity of *Plasmodium vivax*. *Memórias do Instituto Oswaldo Cruz*, 106, 12-26.
- Canier, L., Khim, N., Kim, S., Sluydts, V., Heng, S., Dourng, D., Eam, R., Chy, S., Khean, C., Loch, K. and Ken, M. (2013). An innovative tool for moving malaria PCR detection of parasite reservoir into the field. *Malaria Journal*, 12(405). doi:10.1186/1475-2875-12-405
- Carlton, J. M., Das, A., and Escalante, A. A. (2013). Genomics, population genetics and evolutionary history of *Plasmodium vivax*. *Advances in Parasitology*, 81, 203-222.
- Carter, R., and Mendis, K. N. (2002). Evolutionary and historical aspects of the burden of malaria. *Clinical Microbiology Reviews*. 15(4), 564–594. doi: 10.1128/CMR.15.4.564-594.2002.
- Cavasini, C. E., de Mattos, L. C., Couto, Á. A. D. A., Bonini-Domingos, C. R., Valencia, S. H., de Souza Neiras, W. C., Alves, R. T., Rossit, A. R. B., Castilho, L. and Machado, R. L. D. (2007). *Plasmodium vivax* infection among Duffy antigen-negative individuals from the Brazilian Amazon region: an exception? *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 101(10), 1042-1044. doi:10.1016/j.trstmh.2007.04.011
- Cawthon Lang, K. A. (2006). Primate Factsheets: Long-tailed macaque (*Macaca fascicularis*) Taxonomy, Morphology, & Ecology. Retrieved from: [http://pin.primate.wisc.edu/factsheets/entry/long-tailed\\_macaque](http://pin.primate.wisc.edu/factsheets/entry/long-tailed_macaque)

Centers for Disease Control and Prevention. (2014). Malaria Facts. Atlanta, U.S.A.  
Retrieved from: <http://www.cdc.gov/malaria/about/biology/>

Chang, M. S., Hii, J., Buttner, P., and Mansoor, F. (1997). Changes in abundance and behaviour of vector mosquitoes induced by land use during the development of an oil palm plantation in Sarawak. *Transaction of the Royal Society of Tropical Medicine and Hygiene*, 91(4), 382–386. doi:10.1016/S0035-9203(97)90248-0

Cheong, F. W., Lau, Y. L., Fong, M. Y., and Mahmud, R. (2013). Evaluation of recombinant *Plasmodium knowlesi* merozoite surface protein-133 for detection of human malaria. *The American Journal of Tropical Medicine and Hygiene*, 88(5), 835-840. doi:10.4269/ajtmh.12-0250

Cheong, F. W., Fong, M. Y., Lau, Y. L., and Mahmud, R. (2013). Immunogenicity of bacterial-expressed recombinant *Plasmodium knowlesi* merozoite surface protein-142 (MSP-142). *Malaria Journal*, 12, 454.

Cheong, W. H., Warren, M. W., Omar, A. H., and Mahadevan, S. (1965). *Anopheles balabacensis balabacensis* identified as vector of simian malaria in Malaysia. *Science* 150(3701), 1314-5.

Chew, C. H., Lim, Y. A. L., Lee, P. C., Mahmud, R., and Chua, K. H. (2012). Hexaplex PCR detection system for identification of five human *Plasmodium* species with an internal control. *Journal of Clinical Microbiology*, 50(12), 4012. doi: 10.1128/JCM.06454-11.

Chilton, D., Malik, A. N., Armstrong, M., Kettelhut, M., Parker-Williams, J., and Chiodini, P. L. (2006). Use of rapid diagnostic tests for diagnosis of malaria in the UK. *Journal of Clinical Pathology*, 59(8), 862–866. doi: 10.1136/jcp.2005.032904

Chin, W., Contacos, P. G., Coatney, G. R., and Kimball, H. R. (1965). A naturally acquired quotidian-type malaria in man transferrable to monkeys. *Science* 149(3686), 865. doi: 10.1126/science.149.3686.865

Chin, W., Contacos, P. G., Collins, W. E., Jeter, M. H., and Alpert, E. (1968). Experimental mosquito-transmission of *Plasmodium knowlesi* to man and monkey. *The American Journal of Tropical Medicine and Hygiene*, 17(3), 355–358.

Choi, Y. K., Choi, K. M., Park, M. H., Lee, E. G., Kim, Y. J., Lee, B. C., Cho, S. H., Rhie, H. G., Lee, H. S., Yu, J. R. and Lee, J. S., (2010). Rapid dissemination of newly introduced *Plasmodium vivax* genotypes in South Korea. *The American journal of tropical medicine and hygiene*, 82(3), 426-432.

Chotivanich, K., Silamut, K., and Day, N. P. (2007). Laboratory diagnosis of malaria infection-A short review of methods. *New Zealand Journal of Medical Laboratory Science*, 61(1), 4.

- Chou, M., Kim, S., Khim, N., Chy, S., Sum, S., Dourng, D., Canier, L., Nguon, C., and Menard, D. (2012). Performance of “VIKIA Malaria Ag Pf/Pan” (IMACCESS®), a new malaria rapid diagnostic test for detection of symptomatic malaria infections. *Malaria Journal*, 11(295). doi: 10.1186/1475-2875-11-295.
- Coatney, G. R., Elder, H. A., Contacos, P. G., Getz, M. E., Greenland, R., Rossan, R. N., and Schmidt, L. H. (1961). Transmission of the M strain of *Plasmodium cynomolgi* to man [Abstract]. *The American Journal of Tropical Medicine and Hygiene*, 10, 673–678.
- Coatney, G. R., Chin, W., Contacos, P. G., and King, H. K. (1966). *Plasmodium inui*, a quartan-type malaria parasite of Old World monkeys transmissible to man. *The Journal of Parasitology*. 52(4), 660–663.
- Coatney, G. R., Collins, W. E., Warren, M., and Contacos, P. G. (1971). The Primate Malaria. Washington D.C.: U.S. Government Printing Office.
- Cogswell, F. B. (1992). The hypnozoite and relapse in primate malaria. *Clinical Microbiology Reviews*, 5(1), 26-35.
- Cogswell, F. B. (2000). Malaria and piroplasm of non-human primates. In: D. D. Bowman (Ed.), *Companion and Exotic Animal Parasitology*. Covington, LA: International Veterinary Information Service ([www.ivis.org](http://www.ivis.org)). Available from: [http://www.ivis.org/advances/Parasit\\_Bowman/cogswell\\_primate/ivis.pdf](http://www.ivis.org/advances/Parasit_Bowman/cogswell_primate/ivis.pdf).
- Collins, W. E., Contacos, P. G., and Guinn, E. G. (1967). Studies on the transmission of simian malarias II. Transmission of the H strain of *Plasmodium knowlesi* by *Anopheles balabacensis balabacensis*. *Journal of Parasitology*, 53(4), 841-844.
- Collins, W. E., Contacos, P. G., Skinner, J. C., and Guinn, E. G. (1971). Studies on the transmission of simian malarias. IV. Further studies on the transmission of *Plasmodium knowlesi* by *Anopheles balabacensis balabacensis* mosquitoes. *Journal of Parasitology* 57(5), 961-966.
- Collins, W. E., and Barnwell, J. W. (2009). *Plasmodium knowlesi*: finally being recognized. *Journal of Infectious Diseases*, 199(8), 1107-1108. doi: 10.1086/597415
- Collins, W. E. (2012). *Plasmodium knowlesi*: A malaria parasite of monkeys and humans. *Annual Review of Entomology*, 57(1), 107–121. doi: 10.1146/annurev-ento-121510-133540
- Connelly, C. R., Baker, R. H., Morris, C. D., and Naya, J. K. (1990). Human Malaria. *SP134: Pests in and around the Florida Home*.
- Contacos, P. G., Lunn, J. S., Coatney, G. R., Kilpatrick, J. W., and Jones, F. E. (1963). Quartan-type malaria parasite of new world monkeys transmissible to man. *Science*, 142(3593), 676.

- Contacos, P. G., Coatney, G. R., Orihel, T. C., Collins, W. E., Chin, W., and Jeter, M. H. (1970). Transmission of *Plasmodium schwetzi* from the chimpanzee to man by mosquito bite [Abstract]. *The American Journal of Tropical Medicine and Hygiene*, 19(2), 190–195.
- Conway, D. J., Cavanagh, D. R., Tanabe, K., Roper, C., Mikes, Z. S., Sakihama, N., Bojang, K. A., Oduola, A. M., Kremsner, P. G., Arnot, D. E. and Greenwood, B. M., (2000). A principal target of human immunity to malaria identified by molecular population genetic and immunological analyses. *Nature Medicine*, 6(6), 689-692.
- Conway, D. J. (2007). Molecular epidemiology of malaria. *Clinical Microbiology Reviews*, 20(1), 188–204. doi:10.1128/CMR.00021-06
- Cox, F. E. G. (2010). History of the discovery of the malaria parasites and their vectors. *Parasites & Vectors*, 3(1), 5. doi:10.1186/1756-3305-3-5
- Cox-Singh, J. (2009). *Knowlesi* malaria in Vietnam. *Malaria Journal*, 8, 269. doi:10.1186/1475-2875-8-269
- Cox-Singh, J., and Balbir, S. (2008). *Knowlesi* malaria: newly emergent and of public health importance?, *Trends in Parasitology*, 24(9), 406-410.
- Cox-Singh, J., Davis, T. M., Lee, K. S., Shamsul, S. S., Matusop, A., Ratnam, S., Rahman, H. A., Conway, D. J., and Singh, B. (2008). *Plasmodium knowlesi* malaria in humans is widely distributed and potentially life threatening. *Clinical Infectious Disease*, 46(2), 165–171. doi: 10.1086/524888
- Cox-Singh, J., Hiu, J., Lucas, S. B., Divis, P. C., Zulkarnaen, M., Chandran, P., Wong, K.T., Adem, P., Zaki, S.R., Singh, B., and Krishna, S. (2010). Severe malaria—A case of fatal *Plasmodium knowlesi* infection with post-mortem findings: A case report. *Malaria. Journal*, 9(10), 10-1186. doi:10.1186/1475-2875-9-10.
- Cui, L., Escalante, A. A., Imwong, M., and Snounou, G. (2003). The genetic diversity of *Plasmodium vivax* populations. *Trends in Parasitology*, 19(5), 220-226. doi:10.1016/S1471-4922(03)00085-0
- Daneshvar, C., Davis, T. M., Cox-Singh, J., Rafa'ee, M. Z., Zakaria, S. K., Divis, P. C., and Singh, B. (2009). Clinical and laboratory features of human *Plasmodium knowlesi* infection. *Clinical Infectious Disease*, 49(6), 852–860. doi: 10.1086/605439
- Deane, L. M., Deane, M. P., and Ferreira, N. J. (1966). Studies on transmission of simian malaria and on a natural infection of man with *Plasmodium simium* in Brazil. *Bulletin of the World Health Organization*, 35(5), 805–808.
- Desai, M., ter Kuile, F. O., Nosten, F., McGready, R., Asamoah, K., Brabin, B., and Newman, R. D. (2007). Epidemiology and burden of malaria in pregnancy. *The Lancet Infectious Diseases*, 7(2), 93-104.

- De Silva, J. R., Lau, Y. L., & Fong, M. Y. (2014). Genotyping of the duffy blood group among *Plasmodium knowlesi*-infected patients in Malaysia. *PLoS one*, 9(9), e108951. doi:10.1371/journal.pone.0108951.
- Di Giovanni, L., Cochrane, A. H., and Enea, V. (1990). On the evolutionary history of the circumsporozoite protein in *Plasmodia*. *Experimental Parasitology*, 70(4), 373-381. doi:10.1016/0014-4894(90)90120-2
- Divis, P. C., Singh, B., Anderios, F., Hisam, S., Matusop, A., Kocken, C. H., and Conway, D. J. (2015). Admixture in humans of two divergent *Plasmodium knowlesi* populations associated with different macaque host species. doi:10.1371/journal.ppat.1004888
- Dronamraju, K. R., & Arese, P. (Eds.). (2006). *Malaria: genetic and evolutionary aspects*. Springer Science & Business Media.
- Ennis, J. G., Teal, A. E., Habura, A., Madison-Antenucci, S., Keithly, J. S., Arguin, P. M., Hwang, J. (2009). Simian malaria in a US traveler—New York, 2008. *Morbidity and Mortality Weekly Report*, 58(9), 229–232.
- Erdman, L. K., and Kain, K. C. (2008). Molecular diagnostic and surveillance tools for global malaria control. *Travel Medicine and Infectious Disease*, 6(1), 82-99. doi:10.1016/j.tmaid.2007.10.001
- Escalante, A. A., Barrio, E., and Ayala, F. J. (1995). Evolutionary origin of human and primate malarias: evidence from the circumsporozoite protein gene. *Molecular Biology and Evolution*, 12(4), 616-626.
- Escalante, A. A., Lal, A. A., and Ayala, F. J. (1998). Genetic polymorphism and natural selection in the malaria parasite *Plasmodium falciparum*. *Genetics*, 149(1), 189-202.
- Escalante, A. A., Cornejo, O. E., Rojas, A., Udhayakumar, V., and Lal, A. A. (2004). Assessing the effect of natural selection in malaria parasites. *Trends in Parasitology*, 20(8), 388-395.
- Eudey, A. A. (2008). The Crab-Eating macaque (*Macaca fascicularis*): Widespread and rapidly declining. *Primate Conservation*, 23(1), 129-132. doi: <http://dx.doi.org/10.1896/052.023.0115>
- Eyles, D. E., Coatney, G. R., and Getz, M. E. (1960). Vivax-type malaria parasite of macaques transmissible to man. *Science*, 131(3416), 1812-1813.
- Eyles, D. E., Laing, A. B. G., Warren, M., and Sandosham, A. A. (1962). Malaria parasites of the Malayan leaf monkeys of the genus *Presbytis*. *Medical Journal of Malaysia*, 17, 85–86.
- Eyles, D. E., Warren, M., Quinn, E., Wharton, R. H., and Pramachandran, C. (1963). Identification of *Anopheles balabacensis introlatus* as a vectors of monkey malaria in Malaya. *Bulletin of the World Health Organization*, 28(1), 134-5.

- Farrugia, C., Cabaret, O., Botterel, F., Bories, C., Foulet, F., Costa, J. M., and Bretagne, S. (2011). Cytochrome *b* gene quantitative PCR for diagnosing *Plasmodium falciparum* infection in travelers. *Journal of Clinical Microbiology*, 49(6), 2191–2195. doi: 10.1128/JCM.02156-10.
- Felsenstein, J. (1985). Confidence limits on phylogenies: An approach using the bootstrap. *Evolution*, 39, 783–791.
- Figtree, M., Lee, R., Bain, L., Kennedy, T., Mackertich, S., Urban, M., Cheng, Q., and Hudson, B. J. (2010). *Plasmodium knowlesi* in human, Indonesian Borneo. *Emerging Infectious Diseases*, 16(4), 672–674. doi: 10.3201/eid1604.091624
- Fong, Y. L., Cadigan, F. C., and Coatney, G. R. (1971). A presumptive case of naturally occurring *Plasmodium knowlesi* malaria in man in Malaysia. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 65(6), 839–840.
- Fong, M. Y., Lau, Y. L., Chin, L. C., and Al-Mekhlafi, A. M. (2011). Sequence analysis on the mitochondrial COXI gene of recent clinical isolates of *Plasmodium knowlesi* in Klang valley, peninsular Malaysia. *Tropical Biomedicine*, 28(2), 457–463.
- Fong, M. Y., Lau, Y. L., Chang, P. Y., and Anthony, C. N. (2014). Genetic diversity, haplotypes and allele groups of Duffy binding protein (PkDBP $\alpha$ II) of *Plasmodium knowlesi* clinical isolates from Peninsular Malaysia. *Parasites & Vectors*, 7, 161. doi: 10.1186/1756-3305-7-161
- Fooden, J. (1982). Ecogeographic segregation of macaque species. *Primates*, 23(4), 574–579.
- Fooden, J. (1994). Malaria in macaques. *International Journal of Primatology*, 15(4), 573–596.
- Fooden, J. (1995). Systematic review of Southeast Asian long-tail macaques, *Macaca fascicularis* (Raffles, [1821]). *Fieldiana Zoology, new series* (81), 1–206, Chicago, IL: Field Museum of Natural History
- Fooden, J. (2006). Comparative Review of *Fascicularis*-group Species of Macaques (Primates: *Macaca*). *Fieldiana Zoology*, 107, 1–43. doi: [http://dx.doi.org/10.3158/0015-0754\(2006\)107\[1:CROFSM\]2.0.CO;2](http://dx.doi.org/10.3158/0015-0754(2006)107[1:CROFSM]2.0.CO;2)
- Foster, D., Cox-Singh, J., Mohamad, D. S. A., Krishna, S., Chin, P. P., and Singh, B. (2014). Evaluation of three rapid diagnostic tests for the detection of human infections with *Plasmodium knowlesi*. *Malaria Journal*, 13(60), 10–1186.
- Fu, Y. X., and Li, W. H. (1993). Statistical tests of neutrality of mutations. *Genetics* 133, 693–709.
- Galinski, M. R., and Barnwell, W. (2009). Monkey malaria kills four humans. *Trends in Parasitology*, 25(5). doi:10.1016/j.pt.2009.02.002

- Gant, S. M., Clavijo, P., Bai, X., Esko, J. D., and Sinnis, P. (1997). Cell adhesion to a motif shared by the malaria circumsporozoite protein and thrombospondin is mediated by its glycosaminoglycan-binding region and not by CSVTG. *Journal of Biological Chemistry*, 272(31), 19205-19213. doi: 10.1074/jbc.272.31.19205.
- Garske, T., Ferguson, N. M., and Ghani, A. C. (2013). Estimating air temperature and its influence on malaria transmission across Africa. *PLOS ONE*, 8(2), e56487. doi: 10.1371/journal.pone.0056487
- Gasteiger, E., Gattiker, A., Hoogland, C., Ivanyi, I., Appel, R. D., and Bairoch, A. (2003). ExPASy: The proteomics server for in-depth protein knowledge and analysis. *Nucleic Acids Research*, 31 (13), 3784–8. doi:10.1093/nar/gkg563.
- Gething, P. W., Patil, A. P., Smith, D. L., Guerra, C. A., Elyazar, I. R., Johnston, G. L., Tatem, A. J., and Hay, S. I. (2011). A new world malaria map: *Plasmodium falciparum* endemicity in 2010. *Malaria Journal*, 10(378), 1475-2875.
- Godson, G. A., Ellis, J., Svec, P., Schlesinger, D. H., and Nussenzweig, V. (1983). Identification and chemical synthesis of a tandemly repeated immunogenic region of *Plasmodium knowlesi* circumsporozoite protein. *Nature*, 305, 29-33. doi:10.1038/305029a0
- Goh, X. T., Lim, Y. A., Vythilingam, I., Chew, C. H., Lee, P. C., Ngui, R., ... and Chua, K. H. (2013). Increased detection of *Plasmodium knowlesi* in Sandakan division, Sabah as revealed by PlasmoNex. *Malaria Journal*, 12, 264. doi:10.1186/1475-2875-12-264
- Gonzalez, J. M., Hurtado, S., Arevalo-Herrera, M., and Herrera, S. (2001). Variants of the *Plasmodium vivax* circumsporozoite protein (VK210 and VK247) in Colombian isolates. *Memórias do Instituto Oswaldo Cruz*, 96(5), 709-712.
- Greenwood, B. M., and Targett, G. A. T. (2011). Malaria vaccines and the new malaria agenda. *Clinical Microbiology and Infection*, 17(11), 1600-1607. doi: 10.1111/j.1469-0691.2011.03612.x
- Grigg, M. J., William, T., Drakeley, C. J., Jelip, J., von Seidlein, L., Barber, B. E., Fornace, K. M., Anstey, N. M., Yeo, T. W. and Cox, J. (2014). Factors that are associated with the risk of acquiring *Plasmodium knowlesi* malaria in Sabah, Malaysia: a case-control study protocol. *BMJ open*, 4(8), e006004. doi:10.1136/bmjopen-2014-006004
- Guerra, C. A., Gikandi, P. W., Tatem, A. J., Noor, A. M., Smith, D. L., Hay, S. I., and Snow, R. W. (2008). The limits and intensity of *Plasmodium falciparum* transmission: implications for malaria control and elimination worldwide, *PLoS Medicine*, 5(2), e38. doi:10.1371/journal.pmed.0050038

- Guerra, C. A., Howes, R. E., Patil, A. P., Gething, P. W., Van Boeckel, T. P., Temperley, W. H., Kabaria, C. W., Tatem, A. J., Manh, B. H., Elyazar, I. R., and Baird, J. K. (2010). The international limits and population at risk of *Plasmodium vivax* transmission in 2009. *PLoS Neglected Tropical Diseases*, 4(8), e774. doi:10.1371/journal.pntd.0000774
- Gupta, B., Gupta, P., Sharma, A., Singh, V., Dash, A. P., and Das, A. (2010). High proportion of mixed-species *Plasmodium* infections in India revealed by PCR diagnostic assay. *Tropical Medicine & International Health*, 15(7), 819-824. doi:10.1111/j.1365-3156.2010.02549.x
- Haanshuus, C. G., Mohn, S. C., Mørch, K., Langeland, N., Blomberg, B., and Hanevik, K. (2013). A novel, single-amplification PCR targeting mitochondrial genome highly sensitive and specific in diagnosing malaria among returned travellers in Bergen, Norway. *Malaria Journal*, 12(1), 26. doi:10.1186/1475-2875-12-26
- Hafiz, H., and Nor Rasidah, H. (2012). Spatial and temporal distribution of malaria in Peninsular Malaysia from 1998-2010. *Health and the Environment Journal*, 3(3), 46-50.
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. In *Nucleic acids symposium series*, 41, 95-98.
- Hambali, K., Ismail, A., and Md-Zain, B. M. (2012). Daily activity budget of Long-tailed macaques (*Macaca fascicularis*) in Kuala Selangor Nature Park. *International Journal of Basic and Applied Sciences*, 12(4), 47-52.
- Hamilton, A. J., Suhrbier, A., Nicholas, J., and Sinden, R. E. (1988). Immunoelectron microscopic localization of circumsporozoite antigen in the differentiating exoerythrocytic trophozoite of *Plasmodium berghei*. *Cell Biology International Reports*, 12(2), 123-129. doi:10.1016/0309-1651(88)90126-9
- Han, E. T., Watanabe, R., Sattabongkot, J., Khuntirat, B., Sirichaisinthop, J., Iriko, H., Jin, L., Takeo, S., and Tsuboi, T. (2007). Detection of four *Plasmodium* species by genus- and species-specific loop-mediated isothermal amplification for clinical diagnosis. *Journal of Clinical Microbiology*, 45(8), 2521–2528. doi:10.1128/JCM.02117-06
- Hanscheid, T., and Grobusch, M. P. (2002). How useful is PCR in the diagnosis of malaria? *Trends in Parasitology*, 18(9), 395-398. doi:10.1016/S1471-4922(02)02348-6
- Harbach, R. E. (2013). The phylogeny and classification of *Anopheles*. In S. Manguin (Ed.), *Anopheles mosquitoes - New insights into malaria vectors*. ISBN: 978-953-51-1188-7, InTech, doi: 10.5772/54695. Retrieved from: <http://www.intechopen.com/books/anopheles-mosquitoes-new-insights-into-malaria-vectors/the-phylogeny-and-classification-of-anopheles>
- Hasan, A. U., Suguri, S., Sattabongkot, J., Fujimoto, C., Amakawa, M., Harada, M., and Ohmae, H. (2009). Implementation of a novel PCR based method for

detecting malaria parasites from naturally infected mosquitoes in Papua New Guinea. *Malaria Journal*, 8(1), 182. doi:10.1186/1475-2875-8-182

Hay, S. I., Sinka, M. E., Okara, R. M., Kabaria, C. W., Mbithi, P. M., Tago, C. C., Benz, D., Gething, P. W., Howes, R. E., Patil, A. P., and Temperley, W. H. (2010). Developing global maps of the dominant *Anopheles* vectors of human malaria. *PLoS Med.* 7(2):e1000209. doi: 10.1371/journal.pmed.1000209

Hay, S. I., Okiro, E. A., Gething, P. W., Patil, A. P., Tatem, A. J., Guerra, C. A. and Snow, R. W. (2010). Estimating the global clinical burden of *Plasmodium falciparum* malaria in 2007. *PLoS Medicine*, 7(6), e1000290. doi:10.1371/journal.pmed.1000290

Haynes, J. D., Dalton, J. P., Klotz, F. W., McGinniss, M. H., Hadley, T. J., Hudson, D. E., and Miller, L. H. (1988). Receptor-like specificity of a *Plasmodium knowlesi* malarial protein that binds to Duffy antigen ligands on erythrocytes. *The Journal of Experimental Medicine*, 167(6), 1873-1881.

Hoosen, A., and Shaw, M. T. (2011). *Plasmodium knowlesi* in a traveller returning to New Zealand. *Travel Medicine and Infectious Disease*, 9(3), 144–148. doi:10.1016/j.tmaid.2011.03.002

Huang, C. C., Chiang, Y. C., Ji, D. D., Teng, H. J., Liao, M. H., Chang, C. D., and Wu, Y. H. (2010). Molecular cloning and characterization of the circumsporozoite protein gene of *Plasmodium inui* isolated from Formosan macaques (*Macaca cyclopis*) in Taiwan. *Journal of Parasitology*, 96(6), 1145-1151. doi: <http://dx.doi.org/10.1645/GE-2485.1>

Hughes, A. L. (1991). Circumsporozoite protein genes of malaria parasites (*Plasmodium* spp.): evidence for positive selection on immunogenic regions. *Genetics*, 127(2), 345-353.

Hughes, A. L. (1992). Positive selection and interallelic recombination at the merozoite surface antigen-1 (MSA-1) locus of *Plasmodium falciparum*. *Molecular Biology and Evolution*, 9(3), 381-393.

Hulden, L., and Hulden, L. (2011). Activation of the hypnozoite: a part of *Plasmodium vivax* life cycle and survival. *Malaria Journal*, 10(1), 90. doi:10.1186/1475-2875-10-90

Imwong, M., Tanomsing, N., Pukrittayakamee, S., Day, N. P., White, N. J., and Snounou, G. (2009). Spurious amplification of a *Plasmodium vivax* small-subunit RNA gene by use of primers currently used to detect *P. knowlesi*. *Journal of Clinical Microbiology*, 47(12). doi: 10.1128/JCM.00811-09

Irene, L. M. (2011). *Identification and molecular characterisation of simian malaria parasites in wild monkeys of Singapore* (Doctoral dissertation).

- Jeslyn, W. P. S., Huat, T. C., Vernon, L., Irene, L. M. Z., Sung, L. K., Jarrod, L. P., Singh, B., and Ching, N. L. (2011). Molecular epidemiological investigation of *Plasmodium knowlesi* in humans and macaques in Singapore. *Vector-Borne and Zoonotic Diseases*, 11(2), 131-135. doi: 10.1089/vbz.2010.0024
- Jiang, N., Chang, Q., Sun, X., Lu, H., Yin, J., Zhang, Z., Wahlgren, M., and Chen, Q. (2010). Co-infections with *Plasmodium knowlesi* and other malaria parasites, Myanmar. *Emerging Infectious Diseases*, 16(9), 1476-1478. doi: 10.3201/eid1609.100339
- Jiram, A. I., Vythilingam, I., NoorAzian, Y. M., Yusof, Y. M., Azahari, A. H., and Fong, M. Y. (2012). Entomologic investigation of *Plasmodium knowlesi* vectors in Kuala Lipis, Pahang, Malaysia. *Malaria Journal*, 11(1), 213. doi:10.1186/1475-2875-11-213.
- Jongwutiwes, S., Putaporntip, C., Iwasaki, T., Sata, T., and Kanbara, H. (2004). Naturally acquired *Plasmodium knowlesi* malaria in human, Thailand. *Emerging Infectious Diseases*, 10(12), 2211–2213. doi: 10.3201/eid1012.040293
- Jongwutiwes, S., Buppan, P., Kosuvin, R., Seethamchai, S., Pattanawong, U., Sirichaisinthop, J., and Putaporntip, C. (2011). *Plasmodium knowlesi* malaria in humans and macaques, Thailand. *Emerging Infectious Diseases*, 17(10), 1799 –1806. doi: 10.3201/eid1710.110349
- Joveen-Neoh, W. F., Chong, K. L., Wong, C. M., and Lau, T. Y. (2011). Incidence of malaria in the interior division of Sabah, Malaysian Borneo, based on nested PCR. *Journal of Parasitology Research*, 104284. doi:10.1155/2011/104284.
- Joy, D. A., Feng, X., Mu, J., Furuya, T., Chotivanich, K., Krettli, A. U., Ho, M., Wang, A., White, N. J., Suh, E., and Beerli, P. (2003). Early origin and recent expansion of *Plasmodium falciparum*. *Science*, 300(5617), 318–321. doi: 10.1126/science.1081449
- Kantele, A., Marti, H., Felger, I., Müller, D., and Jokiranta, T. S. (2008). Monkey malaria in a European traveler returning from Malaysia. *Emerging Infectious Diseases*, 14(9), 1434 –1436. doi: 10.3201/eid1409.080170
- Kantele, A., and Jokiranta, T. S. (2011). Review of cases with the emerging fifth human malaria parasite, *Plasmodium knowlesi*. *Clinical Infectious Diseases*, 52(11), 1356-1362. doi: 10.1093/cid/cir180
- Kappe, S. H., Buscaglia, C. A., and Nussenzweig, V. (2004). *Plasmodium* sporozoite molecular cell biology. *Annual Review of Cell Developmental Biology*, 20, 29-59. doi: 10.1146/annurev.cellbio.20.011603.150935
- Khim, N., Siv, S., Kim, S., Mueller, T., Fleischmann, E., Singh, B., Divis, P. C. S., Steenkeste, N., Duval, L., Bouchier, C., and Duong, S. (2011). *Plasmodium knowlesi* infection in humans, Cambodia, 2007-2010. *Emerging Infectious Diseases*, 17(10), 1900 –1902. doi: 10.3201/eid1710.110355

- Kho, W. G., Park, Y. H., Chung, J. Y., Kim, J. P., Hong, S. T., Lee, W. J., Kim, T. S., and Lee, J. S. (1999). Two new genotypes of *Plasmodium vivax* circumsporozoite protein found in the Republic of Korea. *The Korean Journal of Parasitology*, 37(4), 265-270.
- Kho, W. G., Chung, J. Y., Sim, E. J., Kim, M. Y., Kim, D. W., Jongwutiwes, S., and Tanabe, K. (2003). A multiplex polymerase chain reaction for a differential diagnosis of *Plasmodium falciparum* and *Plasmodium vivax*. *Parasitology International*, 52(3), 229–236. doi:10.1016/S1383-5769(03)00028-X
- Kimura, M. (1980). A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *Journal of Molecular Evolution*, 16, 111-120.
- Kolakovich, K. A., Ssengoba, A., Wojcik, K., Tsuboi, T., Al-Yaman, F., Alpers, M., and Adams, J. H. (1996). *Plasmodium vivax*: favored gene frequencies of the merozoite surface protein-1 and the multiplicity of infection in a malaria endemic region. *Experimental Parasitology*, 83(1), 11-18.
- Kuo, M. C., Chiang, T. Y., Chan, C. W., Tsai, W. S., and Ji, D. D. (2009). A case report of simian malaria *Plasmodium knowlesi*, in a Taiwanese traveller from Palawan Island. *Taiwan Epidemiology Bulletin*, 25(3), 178 –191.
- Lambrecht, F. L., Dunn, F. L., and Eyles, D. E. (1961). Isolation of *Plasmodium knowlesi* from Philippine macaques. *Nature*, 191, 1117–1118. doi:10.1038/1911117a0
- Lau, T. Y., Joveen-Neoh, W. F., and Chong, K. L. (2011). High incidence of *Plasmodium knowlesi* infection in the interior division of Sabah, Malaysian Borneo. *International Journal of Bioscience, Biochemistry and Bioinformatics*, 1(2), 163-167.
- Lau, T. Y., Joveen-Neoh, W. F., and Chong, K. L. (2011). Molecular detection of *Plasmodium knowlesi* in the interior division of Sabah, Malaysian Borneo. *International Proceedings of Chemical, Biological and Environmental Engineering* (vol.9), 135-139.
- Lau, Y. L., Tan, L. H., Chin, L. C., Fong, M. Y., Noraishah, M. A., and Rohela, M. (2011). *Plasmodium knowlesi* reinfection in human. *Emerging Infectious Diseases*, 17(7), 1314– 1315. doi: 10.3201/eid1707.101295
- Lawler, J., and Hynes, R. O. (1986). The structure of human thrombospondin, an adhesive glycoprotein with multiple calcium-binding sites and homologies with several different proteins. *Journal of Cell Biology*, 103(5), 1635-1648.
- Leclerc, M. C., Hugot, J. P., Durand, P., and Renaud, F. (2004). Evolutionary relationships between 15 Plasmodium species from New and Old World primates (including humans): a 18S rDNA cladistic analysis. *Parasitology*, 129(06), 677-684.

- Lee, C. E., Adeeba, K., and Freigang, G. (2010). Human *Plasmodium knowlesi* infections in Klang Valley, Peninsula Malaysia: A case series. *Medical Journal of Malaysia*, 65(1), 63– 65.
- Lee, K. S., Cox-Singh, J., and Singh, B. (2009). Morphological features and differential counts of *Plasmodium knowlesi* parasites in naturally acquired human infections. *Malaria Journal*, 8, 73. doi:10.1186/1475-2875-8-73
- Lee, K. S., Cox-Singh, J., Brooke, G., Matusop, A., and Singh, B. (2009). *Plasmodium knowlesi* from archival blood films: Further evidence that human infections are widely distributed and not newly emergent in Malaysian Borneo. *International Journal of Parasitology*, 39(10), 1125–1128. doi: 10.1016/j.ijpara.2009.03.003
- Lee, K. S., Divis, P. C., Zakaria, S. K., Matusop, A., Julin, R. A., Conway, D. J., Cox-Singh, J., and Singh, B. (2011). *Plasmodium knowlesi*: reservoir hosts and tracking the emergence in humans and macaques. *PLoS Pathogens*, 7(4), e1002015. doi: 10.1371/journal.ppat.1002015
- Lee, K. S., and Vythilingam, I. (2013). *Plasmodium knowlesi*: Emergent Human Malaria in Southeast Asia. In *Parasites and their vectors* (pp. 5-31). Springer Vienna.
- Lee, W. C., Chin, P. W., Lau, Y. L., Chin, L. C., Fong, M. Y., Yap, C. J., ... and Mahmud, R. (2013). Hyperparasitaemic human *Plasmodium knowlesi* infection with atypical morphology in peninsular Malaysia. *Malaria Journal*, 12(88), 10-1186. doi:10.1186/1475-2875-12-88
- Link, L., Bart, A., Verhaar, N., van Gool, T., Pronk, M., and Scharnhorst, V. (2012). Molecular diagnosis of *Plasmodium knowlesi* in a Dutch traveler by realtime PCR. *Journal of Clinical Microbiology*, 50(7), 2523–2524. doi: 10.1128/JCM.06859-11.
- Lourenço-de-Oliveira, R., and Luz, S. L. (1996). Simian malaria at two sites in the Brazilian Amazon-II: Vertical distribution and frequency of anopheline species inside and outside the forest. *Memórias do Instituto Oswaldo Cruz*, 91(6), 687-694.
- Luchavez, J., Espino, F. E., Curameng, P., Espina, R., Bell, D., Chiodini, P., Nolder, D., Sutherland, C., Lee, K. S., and Singh, B. (2008). Human infections with *Plasmodium knowlesi*, the Philippines. *Emerging Infectious Diseases*, 14(5), 811– 813. doi: 10.3201/eid1405.071407
- Lucchi, N. W., Poorak, M., Oberstaller, J., DeBarry, J., Srinivasamoorthy, G., Goldman, I., Xayavong, M., Da Silva, A. J., Peterson, D. S., Barnwell, J. W., and Kissinger, J., (2012). A new single-step PCR assay for the detection of the zoonotic malaria parasite *Plasmodium knowlesi*. *PLoS One*, 7(2), e31848.
- Luke, T. C., and Hoffman, S. L. (2003). Rationale and plans for developing a non-replicating, metabolically active, radiation-attenuated *Plasmodium falciparum* sporozoite vaccine. *Journal of Experimental Biology*, 206(21), 3803-3808.

- Malaivijitnond, S. U. C. H. I. N. D. A., and Hamada, Y. U. Z. U. R. U. (2008). Current situation and status of long-tailed macaques (*Macaca fascicularis*) in Thailand. *Natural History Journal of Chulalongkorn University*, 8, 185-204.
- Manguin, S., Garros, C., Dusfour, I., Harbach, R. E., and Coosemans, M. (2008). Bionomics, taxonomy, and distribution of the major malaria vector taxa of *Anopheles* subgenus *Cellia* in Southeast Asia: an updated review. *Infection, Genetics and Evolution*, 8(4), 489-503. doi: 10.1016/j.meegid.2007.11.004
- Marchand, R. P., Culleton, R., Maeno, Y., Quang, N. T., and Nakazawa, S. (2011). Co-infections of *Plasmodium knowlesi*, *P. falciparum*, and *P. vivax* among humans and *Anopheles dirus* mosquitoes, Southern Vietnam. *Emerging Infectious Diseases*, 17(7). doi: 10.3201/eid1707.101551
- Marchand, R. P. (2005). The Khanh Phu malaria research project: an overview 1994–2004. *Anopheles dirus* mosquitoes, southern Vietnam. *Emerging Infectious Diseases*, 17, 1232–1239. Retrieved from: [http://www.mcnv.nl/uploads/media/Malaria\\_overview\\_2005\\_18.pdf](http://www.mcnv.nl/uploads/media/Malaria_overview_2005_18.pdf)
- Martin, M. J., Rayner, J. C., Gagneux, P., Barnwell, J. W., and Varki, A. (2005). Evolution of human-chimpanzee differences in malaria susceptibility: relationship to human genetic loss of *N*-glycolylneuraminic acid. *Proceedings of the National Academy of Sciences of the United States of America*, 102(36), 12819–12824.
- McCutchan, T. F., Kissinger, J. C., Touray, M. G., Rogers, M. J., Li, J., Sullivan, M., Braga, E. M., Krettli, A. U., and Miller, L. H. (1996). Comparison of circumsporozoite proteins from avian and mammalian malarias: biological and phylogenetic implications. *Proceedings of the National Academy of Sciences*, 93(21), 11889-11894.
- Ménard, D., Barnadas, C., Bouchier, C., Henry-Halldin, C., Gray, L. R., Ratsimbasona, A., Thonier, V., Carod, J. F., Domarle, O., Colin, Y., and Bertrand, O. (2010). *Plasmodium vivax* clinical malaria is commonly observed in Duffy-negative Malagasy people. *Proceedings of the National Academy of Sciences of the United States of America*, 107(13), 5967-71. doi: 10.1073/pnas.0912496107.
- Mendes, C., Dias, F., Figueiredo, J., Mora, V. G., Cano, J., de Sousa, B., Do Rosário, V. E., Benito, A., Berzosa, P., and Arez, A. P. (2011). Duffy negative antigen is no longer a barrier to *Plasmodium vivax* – Molecular evidences from the African West Coast (Angola and Equatorial Guinea). *PLoS Neglected Tropical Disease*, 5(6), e1192. doi: 10.1371/journal.pntd.0001192.
- Mens, P. F., van Amerongen, A., Sawa, P., Kager, P. A., and Schallig, H. D. (2008). Molecular diagnosis of malaria in the field: Development of a novel 1-step nucleic acid lateral flow immunoassay for the detection of all 4 human *Plasmodium* spp. and its evaluation in Mbita, Kenya. *Diagnostic Microbiology and Infectious Disease*, 61(4), 421-427.

- Miller, L. H., Mason, S. J., Clyde, D. F., and McGinniss, M. H. (1976). The resistance factor to *Plasmodium vivax* in blacks: the Duffy-blood-group genotype, FyFy. [Abstract]. *The New England Journal of Medicine*, 295(6), 302–4. doi:10.1056/NEJM197608052950602. PMID 778616.
- Ministry of Health Malaysia. (2012). Annual Report 2012. 326pp.
- Mohamed, Z., and Roshan, T. M. (2009). Human *Plasmodium knowlesi*: An emerging infection presented with severe thrombocytopenia. *International Medical Journal*, 16(4), 307-310.
- Moody, A. (2002). Rapid Diagnostic Tests for Malaria Parasites. *Clinical Microbiology Reviews*, 15(1), 66-78. doi: 10.1128/CMR.15.1.66-78.2002
- Moorthy, V. S., Good, M. F., and Hill, A. V. (2004). Malaria vaccine developments. *The Lancet*, 363(9403), 150-156. doi:10.1016/S0140-6736(03)15267-1
- Moran, P., and Caras, I. W. (1994). Requirements for glycosylphosphatidylinositol attachment are similar but not identical in mammalian cells and parasitic protozoa. *The Journal of Cell Biology*, 125(2), 333-343. doi: 10.1083/jcb.125.2.333
- Morgan, K., Somboon, P., and Walton, C. (2013). Understanding *Anopheles* diversity in Southeast Asia and its applications for malaria control. In S. Manguin (Ed.), *Anopheles mosquitoes - New insights into malaria vectors*. ISBN: 978-953-51-1188-7, InTech, DOI: 10.5772/55709. Retrieved from: <http://www.intechopen.com/books/anopheles-mosquitoes-new-insights-into-malaria-vectors/understanding-anopheles-diversity-in-southeast-asia-and-its-applications-for-malaria-control>
- Mouatcho, J. C., and Goldring, J. D. (2013). Malaria rapid diagnostic tests: challenges and prospects. *Journal of Medical Microbiology*, 62(10), 1491-1505.
- Moyes, C. L., Henry, A. J., Golding, N., Huang, Z., Singh, B., Baird, J. K., Newton, P. N., Huffman, M., Duda, K. A., Drakeley, C. J., and Elyazar, I. R. (2014). Defining the geographical range of the *Plasmodium knowlesi* reservoir. *PLoS Neglected Tropical Diseases*, 8(3), e2780. doi:10.1371/journal.pntd.0002780
- Mu, J., Joy, D. A., Duan, J., Huang, Y., Carlton, J., Walker, J., Barnwell, J., Beerli, P., Charleston, M. A., Pybus, O. G., and Su, X. Z.. (2005). Host switch leads to emergence of *Plasmodium vivax* malaria in humans. *Molecular Biology and Evolution*, 22(8), 1686-1693.
- Muehlenbein, M. P., Pacheco, M. A., Taylor, J. E., Prall, S. P., Ambu, L., Nathan, S., Alisisto, S., Ramirez, D., and Escalante, A. A., (2015). Accelerated diversification of nonhuman primate malarias in southeast Asia: adaptive radiation or geographic speciation ?. *Molecular Biology and Evolution*, 32(2), 422-439.

- Mueller, I., Galinski, M. R., Baird, J. K., Carlton, J. M., Kocher, D. K., Alonso, P. L., and del Portillo, H. A. (2009). Key gaps in the knowledge of *Plasmodium vivax*, a neglected human malaria parasite. *The Lancet Infectious Diseases*, 9(9), 555-566. doi: 10.1016/S1473-3099(09)70177-X
- Müller, M., and Schlagenhauf, P. (2014). *Plasmodium knowlesi* in travellers, update 2014. *International Journal of Infectious Diseases*, 22, 55-64.
- Murray, C. K., Bell, D., Gasser, R. A., and Wongsrichanalai, C. (2003). Rapid diagnostic testing for malaria. *Tropical Medicine and International Health*, 8(10), 876–883. doi: 10.1046/j.1365-3156.2003.01115.x
- Murray, C. K., Gasser, R. A., Jr, Magill, A. J., and Miller, R. S. (2008). Update on rapid diagnostic testing for malaria. *Clinical Microbiology Reviews*, 21(1), 97–110. doi: 10.1128/CMR.00035-07
- Murray, C. K., and Bennett, J. W. (2009). Rapid diagnosis of malaria. *Interdisciplinary Perspectives on Infectious Diseases*, vol. 2009, Article ID: 415953. doi:10.1155/2009/415953.
- Myung, J. M., Marshall, P., and Sinnis, P. (2004). The *Plasmodium* circumsporozoite protein is involved in mosquito salivary gland invasion by sporozoites. *Molecular and Biochemical Parasitology*, 133(1), 53-59. doi:10.1016/j.molbiopara.2003.09.002
- Nagasawa, H., Procell, P. M., Atkinson, C. T., Campbell, G. H., Collins, W. E., and Aikawa, M. (1987). Localization of circumsporozoite protein of *Plasmodium ovale* in midgut oocysts. *Infection and Immunity*, 55(12), 2928-2932.
- Naing, D. K. S., Anderios, F., and Lin, Z. (2011). Geographic and ethnic distribution of *P. knowlesi* infection in Sabah, Malaysia. *International Journal of Collaborative Research on Internal Medicine & Public Health*, 3(5), 391–400. Retrieved from: <http://iomcworld.com/ijcrimph/ijcrimph-v03-n05-07.htm>
- Nakazawa, S., Marchand, R. P., Quang, N. T., Culleton, R., Manh, N. D., and Maeno, Y. (2009). *Anopheles dirus* co-infection with human and monkey malaria parasites in Vietnam. *International Journal of Parasitology*, 39(14), 1533–1537. doi: 10.3201/eid1707.101551
- Ng, O. T., Ooi, E. E., Lee, C. C., Lee, P. J., Ng, L. C., Pei, S. W., Tu, T. M., Loh, J. P., and Leo, Y. S. (2008). Naturally acquired human *Plasmodium knowlesi* infection, Singapore. *Emerging Infectious Diseases*, 14(5), 814. doi: 10.3201/eid1405.070863
- Nikzad, S., Tan, S. G., Yong S. Y. C., Ng, J., Alitheen, N. B., Khan, R., Rovie-Ryan, J. J., Valdiani, A., Khajaeian, P. and Kanthaswamy, S., (2014). Genetic diversity and population structure of long-tailed macaque (*Macaca fascicularis*) populations in Peninsular Malaysia. *Journal of Medical Primatology*, 43(6), 433-444. doi:10.1111/jmp.12130

- Ninan, T., Nalees, K., Newin, M., Sultan, Q., Than, M. M., Shinde, S., Habana, A. V. F., and Mohd Yusof, N. (2012). *Plasmodium knowlesi* malaria infection in human. *Brunei International Medical Journal*, 8(6), 358-361.
- Nishimoto, Y., Arisue, N., Kawai, S., Escalante, A. A., Horii, T., Tanabe, K., and Hashimoto, T. (2008). Evolution and phylogeny of the heterogeneous cytosolic SSU rRNA genes in the genus *Plasmodium*. *Molecular Phylogenetics and Evolution*, 47(1), 45-53. doi:10.1016/j.ympev.2008.01.031
- Ohrt, C., Purnomo, Sutamihardia, M. A., Tang, D., and Kain, K. C. (2002). Impact of microscopy error on estimates of protective efficacy in malaria prevention trials. *Journal of Infectious Diseases*, 186(4), 540-546. doi: 10.1086/341938.
- Ong, C. W., Lee, S. Y., Koh, W. H., Ooi, E. E., and Tambyah, P. A. (2009). Monkey malaria in humans: a diagnostic dilemma with conflicting laboratory data. *The American Journal of Tropical Medicine and Hygiene*, 80(6), 927-928.
- Ozaki, L. S., Svec, P., Nussenzweig, R. S., Nussenzweig, V., and Godson, G. N. (1983). Structure of the *Plasmodium knowlesi* gene coding for the circumsporozoite protein. *Cell*, 34(3), 815-822.
- Padley, D., Moody, A. H., Chiodini, P. L., and Saldanha, J. (2003). Use of rapid, single-round, multiplex PCR to detect malarial parasites and identify the species present. *Annals of Tropical Medicine and Parasitology*, 97(2), 131-137.
- Pain, A., Böhme, U., Berry, A. E., Mungall, K., Finn, R. D., Jackson, A. P., Mourier, T., Mistry, J., Pasini, E. M., Aslett, M. A., and Balasubramanian, S. (2008). The genome of the simian and human malaria parasite *Plasmodium knowlesi*. *Nature*, 455(7214), 799-803. doi:10.1038/nature07306
- Patsoula, E., Spanakos, G., Sofianatou, D., Parara, M., and Vakalis, N. C. (2003). A single-step, PCR-based method for the detection and differentiation of *Plasmodium vivax* and *P. falciparum* [Abstract]. *Annals of Tropical Medicine and Parasitology*, 97(1), 15-21.
- Perandin, F., Manca, N., Calderaro, A., Piccolo, G., Galati, L., Ricci, L., Medici, M.C., Arcangeletti, M. C., Snounou, G., Dettori, G., and Chezzi, C. (2004). Development of a real-time PCR assay for detection of *Plasmodium falciparum*, *Plasmodium vivax*, and *Plasmodium ovale* for routine clinical diagnosis. *Journal of Clinical Microbiology*, 42(3), 1214 -1219. doi: 10.1128/JCM.42.3.1214-1219.2004
- Persson, C., Oliveira, G. A., Sultan, A. A., Bhanot, P., Nussenzweig, V., and Nardin, E. (2002). Cutting edge: A new tool to evaluate human pre-erythrocytic malaria vaccines: rodent parasites bearing a hybrid *Plasmodium falciparum* circumsporozoite protein. *The Journal of Immunology*, 169(12), 6681-6685. doi: 10.4049/jimmunol.169.12.6681

- Pinzon-Ortiz, C., Friedman, J., Esko, J., and Sinnis, P. (2001). The binding of the circumsporozoite protein to cell surface heparan sulfate proteoglycans is required for *Plasmodium* sporozoite attachment to target cells. *Journal of Biological Chemistry*, 276(29), 26784-26791. doi: 10.1074/jbc.M104038200
- Putaporntip, C., Hongsrimuang, T., Seethamchai, S., Kobasa, T., Limkittikul, K., Cui, L., and Jongwutiwes, S. (2009). Differential prevalence of *Plasmodium* infections and cryptic *Plasmodium knowlesi* malaria in humans in Thailand. *Journal of Infectious Diseases*, 199(8), 1143-1150. doi: 10.1086/597414
- Putaporntip, C., Jongwutiwes, S., Thongaree, S., Seethamchai, S., Grynberg, P., and Hughes, A. L. (2010). Ecology of malaria parasites infecting Southeast Asian macaques: evidence from cytochrome b sequences. *Molecular Ecology*, 19(16), 3466-76. doi:10.1111/j.1365-294X.2010.04756.x
- Putaporntip, C., Thongaree, S., and Jongwutiwes, S. (2013). Differential sequence diversity at merozoite surface protein-1 locus of *Plasmodium knowlesi* from humans and macaques in Thailand. *Infection, Genetics and Evolution*, 18, 213-219. doi:10.1016/j.meegid.2013.05.019
- Rajahram, G. S., Barber, B. E., William, T., Menon, J., Anstey, N. M., and Yeo, T. W. (2012). Deaths due to *Plasmodium knowlesi* malaria in Sabah, Malaysia: association with reporting as *Plasmodium malariae* and delayed parenteral artesunate. *Malaria Journal*, 11(1), 284. doi:10.1186/1475-2875-11-284
- Ramasamy, R. (2014). Zoonotic malaria—global overview and research and policy needs. *Frontiers in Public Health*, 2. doi: 10.3389/fpubh.2014.00123
- Ramaswami, A., Pisharam, J. K., Aung, H., Ghazala, K., Maboud, K., Vui, H. C., and Tan, J. (2013). Co-incidental *Plasmodium knowlesi* and Mucormycosis infections presenting with acute kidney injury and lower gastrointestinal bleeding. *American Journal of Case Reports*, 14, 103-105. doi: 10.12659/AJCR.883879
- Rathore, D., Hrstka, S. C. L., Sacci, J. B. Jr., Vega, P. D. I., Linhardt, R. J., Kumar, S., and McCutchan, T. F. (2003). Molecular mechanism of host specificity in *Plasmodium falciparum* infection: Role of circumsporozoite protein. *Journal of Biological Chemistry*, 278(42), 40905-10. doi: 10.1074/jbc.M306250200
- Reid, J. A. (1968). Studies from the Institute for Medical Research Malaysia. *Anopheline Mosquitoes of Malaya and Borneo*. England: Staples Printers Limited.
- Rich, S. M., Licht, M. C., Hudson, R. R., and Ayala, F. J. (1998). Malaria's Eve: evidence of a recent population bottleneck throughout the world populations of *Plasmodium falciparum*. *Proceedings of the National Academy of Sciences*, 95(8), 4425-4430.
- Rich, S. M., and Ayala, F. J. (2006). Evolutionary Origins of Human Malaria Parasites. In K. R. Dronamraju, and P. Arese (Eds.), *Malaria: Genetic and Evolutionary Aspects* (pp. 125-146). New York: Springer.

- Riopel, L. (2013). Background Paper 6.10 Malaria. Retrieved from: [http://cdrwww.who.int/medicines/areas/priority\\_medicines/BP6\\_10malaria.pdf](http://cdrwww.who.int/medicines/areas/priority_medicines/BP6_10malaria.pdf).
- Roll Back Malaria Partnership. (2012). *Progress and Impact Series: Defeating Malaria in Asia, the Pacific, Americas, Middle East and Europe*. Geneva, Switzerland.
- Rougemont, M., Saanen, M. V., Sahli, R., Hinrikson, H. P., Bille, J., and Jaton, K. (2004). Detection of four *Plasmodium* species in blood from human by 18S rRNA gene subunit-based and species-specific realtime PCR assays. *Journal of Clinical Microbiology*, 42(12), 5636–5643. doi: 10.1128/JCM.42.12.5636-5643.2004
- Ryan, J. R., Stoute, J. A., Amon, J., Dunton, R. F., Mtalib, R., Koros, J., ... and Rosenberg, R. (2006). Evidence for transmission of *Plasmodium vivax* among a Duffy antigen negative population in western Kenya. *American Journal of Tropical Medicine and Hygiene*, 75(4), 575–581.
- Sabbatani, S., Fiorino, S., and Manfredi, R. (2010). The emerging of the fifth malaria parasite (*Plasmodium knowlesi*): a public health concern?. *Brazilian Journal of Infectious Diseases*, 14(3), 299-309. doi: <http://dx.doi.org/10.1590/S1413-86702010000300019>
- Saitou, N., and Nei, M. (1987). The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4(4), 406-425.
- Sallum, M. A. M., Peyton, E. L., and Wilkerson, R. C. (2005). Six new species of the *Anopheles leucosphyrus* group, reinterpretation of *An. elegans* and vector implications. *Medical and Veterinary Entomology*, 19(2), 158-199. doi: 10.1111/j.0269-283X.2005.00551.x
- Sallum, M. A. M., Peyton, E. L., Harrison, B. A., and Wilkerson, R. C. (2005). Revision of the *Leucosphyrus* group of *Anopheles* (*Cellia*) (Diptera, Culicidae). *Revista Brasileira de Entomologia*, 49, 01–152.
- Sanders, K. C., Rundi, C., Jelip, J., Rashman, Y., Gueye, C. S., and Gosling, R. D. (2014). Eliminating malaria in Malaysia: the role of partnerships between the public and commercial sectors in Sabah. *Malaria Journal*, 13(1), 24. doi:10.1186/1475-2875-13-24
- Saul, A., Nussenzweig, V., and Nussenzweig, R. S. (2004). Rationale for malaria vaccine development. *Novel Vaccination Strategies*, 479-503. doi: 10.1002/3527601449.ch22
- Scanlon, J. E., Reid, J. A., and Cheong, W. H. (1968). Ecology of *Anopheles* vectors of malaria in the Oriental region. *Cahiers ORSTOM ser Entomology Medicine*, 6, 237–246.

- Schmidt, L. H., Greenland, R., and Genther, C. S. (1961). The transmission of *Plasmodium cynomolgi* to man [Abstract]. *American Journal of Tropical Medicine and Hygiene*, 10:679–688.
- Seethamchai, S., Putaporntip, C., Malaivijitnond, S., Cui, L., and Jongwutiwes, S. (2008). Malaria and *Hepatocystis* species in wild macaques, southern Thailand. *American Journal of Tropical Medicine and Hygiene*, 78(4), 646-653.
- Sermwittayawong, N., Singh, B., Nishibuchi, M., Sawangjaroen, N., and Vuddhakul, V. (2012). Human *Plasmodium knowlesi* infection in Ranong province, southwestern border of Thailand. *Malaria Journal*, 11(1), 36. doi:10.1186/1475-2875-11-36.
- Sharma, S., Svec, P., Mitchell, G. H., and Godson, G. N. (1985). Diversity of circumsporozoite antigen genes from two strains of the malarial parasite *Plasmodium knowlesi*. *Science*, 229(4715), 779-782.
- Sidjanski, S. P., Vanderberg, J. P., and Sinnis, P. (1997). *Anopheles stephensi* salivary glands bear receptors for region I of the circumsporozoite protein of *Plasmodium falciparum*. *Molecular and Biochemical Parasitology*, 90(1), 33-41. doi:10.1016/S0166-6851(97)00124-2
- Singh, A. P., Puri, S. K., and Chitnis, C. E. (2002). Antibodies raised against receptor-binding domain of *Plasmodium knowlesi* Duffy binding protein inhibit erythrocyte invasion. *Molecular and Biochemical Parasitology*, 121(1), 21-31. doi:10.1016/S0166-6851(02)00017-8
- Singh, A. P., Ozwara, H., Kocken, C. H., Puri, S. K., Thomas, A. W., and Chitnis, C. E. (2005). Targeted deletion of *Plasmodium knowlesi* Duffy binding protein confirms its role in junction formation during invasion. *Molecular Microbiology*, 55(6), 1925-1934. doi:10.1111/j.1365-2958.2005.04523.x
- Singh, B. (1997). Molecular methods for diagnosis and epidemiological studies of parasitic infections. *International Journal of Parasitology*, 27(10), 1135-45
- Singh, B., Bobogare, A., Cox-Singh, J., Snounou, G., Abdullah, M. S., and Rahman, H. A. (1999). A genus- and species-specific nested polymerase chain reaction malaria detection assay for epidemiologic studies. *American Journal of Tropical Medicine and Hygiene*, 60(4), 687–692.
- Singh, B., Sung, L. K., Matusop, A., Radhakrishnan, A., Shamsul, S. S., Cox-Singh, J., Thomas, A. and Conway, D. J., (2004). A large focus of naturally acquired *Plasmodium knowlesi* infections in human beings. *The Lancet*, 363(9414), 1017-1024. doi:10.1016/S0140-6736(04)15836-4
- Singh, B., and Cox-Singh, J. (2001). Parasites that cause problems in Malaysia: soil-transmitted helminths and malaria parasites. *TRENDS in Parasitology*, 17(12), 597-600.

- Singh, B., and Daneshvar, C. (2010). *Plasmodium knowlesi* malaria in Malaysia. *Medical Journal of Malaysia* 65(3), 166–172.
- Singh, B., and Daneshvar, C. (2013). Human infections and detection of *Plasmodium knowlesi*. *Clinical Microbiology*, 26(2), 165–184. doi: 10.1128/CMR.00079-12
- Sinka, M. E., Bangs, M. J., Manguin, S., Chareonviriyaphap, T., Patil, A. P., Temperley, W. H. and Hay, S. I. (2011). The dominant *Anopheles* vectors of human malaria in the Asia-Pacific region: occurrence data, distribution maps and bionomic précis. *Parasites & Vectors*, 4(1), 89. doi:10.1186/1756-3305-4-89
- Sinka, M. E., Bangs, M. J., Manguin, S., Rubio-Palis, Y., Chareonviriyaphap, T., Coetzee, M. and Hay, S. I. (2012). A global map of dominant malaria vectors. *Parasites & Vectors*, 5(69). doi:10.1186/1756-3305-5-69
- Sinnis, P., Clavijo, P., Fenyö, D., Chait, B. T., Cerami, C., and Nussenzweig, V. (1994). Structural and functional properties of region II-plus of the malaria circumsporozoite protein. *The Journal of Experimental Medicine*, 180(1), 297–306. doi: 10.1084/jem.180.1.297
- Sinton, J. A., and Mulligan, H. W. (1932). A critical review of the literature relating to the identification of the malaria parasites recorded from monkeys of the families *Cercopithecidae* and *Colobidae*. *Rec. Malar. Surv. India* III:24.
- Sinton, J. A., and Mulligan, H. W. (1933). A critical review of the literature relating to the identification of the malaria parasites recorded from monkeys of the families *Cercopithecidae* and *Colobidae*. *Rec. Malar. Surv. India* III:62.
- Southwick, C. H., and Cadigan Jr, F. C. (1972). Population studies of Malaysian primates. *Primates*, 13(1), 1-18.
- Snounou, G., Viriyakosol, S., Zhu, X. P., Jarra, W., Pinheiro, L., Do Rosario, V. E., Thaithong, S., and Brown, K. N. (1993). High sensitivity of detection of human malaria parasites by the use of nested polymerase chain reaction. *Molecular and Biochemical Parasitology*, 61(2), 315–320. doi: 10.1016/0166-6851(93)90077-B
- Snounou, G., Viriyakosol, S., Jarra, W., Thaithong, S., and Brown, K. N. (1993). Identification of the four human malaria parasite species in field samples by the polymerase chain reaction and detection of a high prevalence of mixed infections. *Molecular and Biochemical Parasitology*, 58(2), 283-292.
- Snounou, G., and Pérignon, J. L. (2013). Malariotherapy: insanity at the service of malariology. *Advances in Parasitology*, 81(6), 223-55.
- Southwick, C. H., and Cadigan Jr, F. C. (1972). Population studies of Malaysian primates. *Primates*, 13(1), 1-18.

- Souza-Neiras, W. C. D., Melo, L. M. S. D., and Machado, R. L. D. (2007). The genetic diversity of *Plasmodium vivax*: a review. *Memórias do Instituto Oswaldo Cruz*, 102(3), 245-254.
- Spangler, W. L., Gribble, D., Abildgaard, C, and Harrison, J. (1978). *Plasmodium knowlesi* Malaria in the Rhesus Monkey. *Veterinary Pathology*, 15(1), 83-91.
- Spitz, A. J. W. (1959). Malaria infection of the placenta and its influence on the incidence of prematurity in eastern Nigeria. *Bulletin of the World Health Organization*, 21(2), 242.
- Steenkeste, N., Incardona, S., Chy, S., Duval, L., Ekala, M. T., Lim, P., Hewitt, S., Sochantha, T., Socheat, D., Rogier, C. and Mercereau-Puijalon, O., (2009). Towards high-throughput molecular detection of *Plasmodium*: new approaches and molecular markers. *Malaria Journal*, 8(1), 86. doi:10.1186/1475-2875-8-86
- Steketee, R. W., Nahlen, B. L., Parise, M. E., and Menendez, C. (2001). The burden of malaria in pregnancy in malaria-endemic areas. *The American Journal of Tropical Medicine and Hygiene*, 64(1), 28-35.
- Stewart, M. J., Nawrot, R. J., Schulman, S., and Vanderberg, J. P. (1986). *Plasmodium berghei* sporozoite invasion is blocked *in vitro* by sporozoite-immobilizing antibodies. *Infection and Immunity*, 51(3), 859-864.
- Sulistyaningsih, E., Fitri, L. E., Löscher, T., and Berens-Riha, N. (2010). Diagnostic difficulties with *Plasmodium knowlesi* infection in humans. *Emerging Infectious Diseases*, 16(6), 1033–1034. doi: 10.3201/eid1606.100022
- Suwonkerd, W., Rithison, W., Ngo, C. T., Tainchum, K., Bangs, M. J., and Chareonviriyaphap, T. (2013). Vector Biology and Malaria Transmission in Southeast Asia. In S. Manquin (Ed.), *Anopheles mosquitoes – New insights into malaria vectors*. ISBN: 978-953-51-1188-7, InTech, doi: 10.5772/56347. Retrieved from <http://www.intechopen.com/books/anopheles-mosquitoes-new-insights-into-malaria-vectors/vector-biology-and-malaria-transmission-in-southeast-asia#article-front>
- Sullivan, J. S., Morris, C. L., Richardson, B. B., Galland, G. G., Sullivan, J. J., and Collins, W. E. (1996). Sporozoite transmission of three strains of *Plasmodium knowlesi* to *Aotus* and *Saimiri* monkeys. *Journal of Parasitology*, 82(2), 268–71
- Swan, H., Sloan, L., Muyombwe, A., Chavalitsewinkoon-Petmitr, P., Krudsood, S., Leowattana, W., and Rosenblatt, J. (2005). Evaluation of a real-time polymerase chain reaction assay for the diagnosis of malaria in patients from Thailand. *American Journal of Tropical Medicine and Hygiene*, 73(5), 850-854.

- Tachibana, S. I., Sullivan, S. A., Kawai, S., Nakamura, S., Kim, H. R., Goto, N., Arisue, N., Palacpac, N. M., Honma, H., Yagi, M. and Tougan, T. (2012). *Plasmodium cynomolgi* genome sequences provide insight into *Plasmodium vivax* and the monkey malaria clade. *Nature Genetics*, 44(9), 1051-1055.
- Tajima, F. (1983). Evolutionary relationship of DNA sequences in finite populations. *Genetics* 105(2), 437–460.
- Takala, S. L., and Plowe, C. V. (2009). Genetic diversity and malaria vaccine design, testing and efficacy: preventing and overcoming ‘vaccine resistant malaria’. *Parasite Immunology*, 31(9), 560-573. doi: 10.1111/j.1365-3024.2009.01138.x
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. (2013). MEGA 6: molecular evolutionary genetics analysis version 6.0. *Molecular Biology and Evolution*, 30(12), 2725-2729.
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., and Kumar, S. (2011). MEGA 5: molecular evolutionary genetics analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution*, 28(10), 2731-2739.
- Tan, C. H., Vytilingam, I., Matusop, A., Chan, S. T., Singh, B. (2008). Bionomics of *Anopheles latens* in Kapit, Sarawak, Malaysian Borneo in relation to the transmission of zoonotic simian malaria parasite *Plasmodium knowlesi*. *Malaria Journal*, 7(52), 10-1186. doi:10.1186/1475-2875-7-52.
- Tangpukdee, N., Duangdee, C., Wilairatana, P., and Krudsood, S. (2009). Malaria diagnosis: A brief review. *Korean Journal of Parasitology*, 47(2), 93-102. doi:10.3347/kjp.2009.47.2.93
- Tang, T. H., Salas, A., Ali-Tammam, M., Martínez, M. C., Lanza, M., Arroyo, E., and Rubio, J. M. (2010). First case of detection of *Plasmodium knowlesi* in Spain by real time PCR in a traveller from Southeast Asia. *Malaria Journal*, 9(1), 219. doi:10.1186/1475-2875-9-219.
- Ta, T. H., Hisam, S., Lanza, M., Jiram, A. I., Ismail, N., and Rubio, J. M. (2014). First case of a naturally acquired human infection with *Plasmodium cynomolgi*. *Malaria Journal*, 13(1), 68. doi:10.1186/1475-2875-13-68
- Tavares, R. G., Staggemeier, R., Borges, A. L. P., Rodrigues, M. T., Castelan, L. A., Vasconcelos, J., Anschau, M. E., and Spalding, S. M. (2011). Molecular techniques for the study and diagnosis of parasite infection. *Journal of Venomous Animals and Toxins including Tropical Diseases*, 17(3), 239-248.
- Tewari, R., Spaccapelo, R., Bistoni, F., Holder, A. A., and Crisanti, A. (2002). Function of Region I and II adhesive motifs of *Plasmodium falciparum* circumsporozoite protein in sporozoite motility and infectivity. *Journal of Biological Chemistry*, 277(49), 47613-47618. doi: 10.1074/jbc.M2084532

- Thathy, V., Fujioka, H., Gantt, S., Nussenzweig, R., Nussenzweig, V., and Ménard, R. (2002). Levels of circumsporozoite protein in the *Plasmodium* oocyst determine sporozoite morphology. *The European Molecular Biology Organization Journal*, 21(7), 1586-1596. doi: 10.1093/emboj/21.7.1586
- Thayer, A. M. (2005). Fighting malaria. *Chemical and Engineering News*, 83(43), 69-82.
- Tilley, L., Dixon, M. W., and Kirk, K. (2011). The *Plasmodium falciparum*-infected red blood cell. *International Journal of Biochemistry and Cell Biology*, 43(6), 839-42. doi:10.1016/j.biocel.2011.03.01
- Tsukamoto, M., and Miyata, A. (1978). Surveys on simian malaria parasites and their vector in Palawan Island, the Philippines. *Tropical Medicine*, 20(1), 39 -50.
- Uneke, C. J. (2007). Impact of placental *Plasmodium falciparum* malaria on pregnancy and perinatal outcome in sub-Saharan Africa: I: introduction to placental malaria. *The Yale Journal of Biology and Medicine*, 80(2), 39.
- Van den Eede, P., Van, H. N., Van Overmeir, C., Vythilingam, I., Duc, T. N., Hung, L. X., ... and Erhart, A. (2009). Human *Plasmodium knowlesi* infections in young children in central Vietnam. *Malaria Journal*, 8(1), 249. doi:10.1186/1475-2875-8-249.
- Van de Peer, Y. (2009). Phylogenetic inference based on distance methods. *The Phylogenetic Handbook*, 142-160.
- Van Hellemond, J. J., Rutten, M., Koelewijn, R., Zeeman, A. M., Verweij, J. J., Wismans, P. J., Kocken, C. H., and van Genderen, P. J. (2009). Human *Plasmodium knowlesi* infection detected by rapid diagnostic tests for malaria. *Emerging Infectious Diseases*, 15(9), 1478 –1480. doi: 10.3201/eid1509.090358
- Van Noordwijk, M. A., and van Schaik, C. P. (1985). Male migration and rank acquisition in wild long-tailed macaques (*Macaca fascicularis*). *Animal Behaviour*, 33(3), 849-861. doi:10.1016/S0003-3472(85)80019-1
- Van Rooyen, C. E., and Pile, G. R. (1935). Observations on infection by *Plasmodium knowlesi* (ape malaria) in the treatment of general paralysis of the insane. *British Medical Journal*, 2(3901), 662.
- Vargas-Serrato, E., Corredor, V., and Galinski, M. R. (2003). Phylogenetic analysis of CSP and MSP-9 gene sequences demonstrates the close relationship of *Plasmodium coatneyi* to *Plasmodium knowlesi*. *Infection, Genetics and Evolution*, 3(1), 67-73. doi:10.1016/S1567-1348(03)00007-8
- Verhulst, N. O., Smallegange, R. C., and Takken, W. (2012). Mosquitoes as potential bridge vectors of malaria parasites from non-human primates to humans. *Frontiers in Physiology*, 3(197). doi: 10.3389/fphys.2012.00197

- Vythilingam, I., Chiang, G. L., Lee, H. L., and Singh, K. I. (1992). Bionomics of important mosquito vectors in Malaysia. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 23(4), 587-603.
- Vythilingam, I., Foo, L. C., Chiang, G. L., Chan, S. T., Eng, K. L., Mahadevan, S., ... and Inder Singh, K. (1995). The impact of permethrin impregnated bednets on the malaria vector *Anopheles maculatus* (Diptera: Culicidae) in aboriginal villages of Pos Betau Pahang, Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health*, 26(2), 354-358.
- Vythilingam, I., Nitiavathy, K., Yi, P., Bakotee, B., Hugo, B., Singh, B., Wirtz, R. A, and Palmer, K. (1999). A highly sensitive, nested polymerase chain reaction based method using simple DNA extraction to detect malaria sporozoites in mosquitos. *Southeast Asian Journal of Tropical Medicine and Public Health*, 30(4), 631-5.
- Vythilingam, I., Chan, S. T., Shanmugratnam, C., Tanrang, H., and Chooi, K. H. (2005). The impact of development and malaria control activities on its vectors in the Kinabatangan area of Sabah, East Malaysia. *Acta Tropica*, 96(1), 24-30. doi:10.1016/j.actatropica.2005.06.022
- Vythilingam, I., Tan, C. H., Asmad, M., Chan, S. T., Lee, K. S., and Singh, B. (2006). Natural transmission of *Plasmodium knowlesi* to humans by *Anopheles latens* in Sarawak, Malaysia. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 100(11), 1087– 1088. 10.1016/j.trstmh.2006.02.006
- Vythilingam, I., Noorazian, Y. M., Huat, T. C., Jiram, A. I., Yusri, Y. M., Aza hari, A. H., ... and LokmanHakim, S. (2008). *Plasmodium knowlesi* in humans, macaques and mosquitoes in Peninsular Malaysia. *Parasites & Vectors*, 1(1), 26. doi:10.1186/1756-3305-1-26
- Vythilingam, I. (2010). Review Paper *Plasmodium knowlesi* in humans: A review on the role of its vectors in Malaysia. *Tropical Biomedicine*, 27(1), 1-12.
- Vythilingam, I. (2012). *Plasmodium knowlesi* and *Wuchereria bancrofti*: Their vectors and challenges for the future. *Frontiers in Physiology*, 3.
- Vythilingam, I., and Hii, J. (2013). Simian malaria parasites: Special emphasis on *Plasmodium knowlesi* and their *Anopheles* vectors in Southeast Asia. In S. Mangnun (Ed.), *Anopheles mosquitoes - New insights into malaria vectors*. ISBN: 978-953-51-1188-7, InTech, doi: 10.5772/54491. Retrieved from: <http://www.intechopen.com/books/anopheles-mosquitoes-new-insights-into-malaria-vectors/simian-malaria-parasites-special-emphasis-on-plasmodium-knowlesi-and-their-anopheles-vectors-in-sout>
- Vythilingam, I., Lim, Y. A., Venugopalan, B., Ngui, R., Leong, C. S., Wong, M. L., Khaw, L., Goh, X., Yap, N., Sulaiman, W. Y. W., and Jeffery, J. (2014). *Plasmodium knowlesi* malaria an emerging public health problem in Hulu Selangor, Selangor, Malaysia (2009-2013): epidemiologic and entomologic analysis. *Parasites & Vectors*, 7(1), 436. doi:10.1186/1756-3305-7-436

- Wagner-Jauregg, J., and Bruetsch, W. L. (1994). The history of the malaria treatment of general paralysis. *The American Journal of Psychiatry*, 151, 231-235.
- Wang, Q., Fujioka, H., and Nussenzweig, V. (2005). Exit of *Plasmodium* sporozoites from oocysts is an active process that involves the circumsporozoite protein. *PLoS Pathogens*, 1(1), e9. doi: 10.1371/journal.ppat.0010009.
- Warhurst, D. C., and Williams, J. E. (1996). Laboratory diagnosis of malaria. *Journal of Clinical Pathology*, 49(7), 533-538.
- Warren, M., Cheong, W. H., Omar, A., and Sandosha. A. A. (1965, January). Ecology of Simian Malaria in Monsoon Forests of Northern Malayan States. In *Journal of Parasitology* (Vol. 51, No. 2 S 2, p. 17). 810 East 10th Street, Lawrence, KS 66044: Amer. Soc. Parasitologists.
- Warren, M. W., and Wharton, R. H. (1963). The vectors of simian malaria: identity, biology, and geographical distribution. *Journal of Parasitology*, 49(6), 892–904.
- Warren, M., Cheong, W. H., Fredericks, H. K., and Coatney, G. R. (1970). Cycles of jungle malaria in West Malaysia. *American Journal of Tropical Medicine and Hygiene*, 19(3), 383–393.
- Weeks, J. R. (2010). Defining urban areas. In *remote sensing of urban and suburban areas* (pp. 33-45). Springer Netherlands.
- Weiss, J. B. (1995). DNA probes and PCR for diagnosis of parasitic infections. *Clinical Microbiology Reviews*, 8(1), 113-130.
- Welch, S. G., McGregor, I. A., and Williams, K. (1977). The Duffy blood group and malaria prevalence in Gambian West Africans [Abstract]. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 71(4), 295–296. doi:10.1016/0035-9203(77)90102-X
- Wharton, R. H., and Eyles, D. E. (1961). *Anopheles hackeri*, a vector of *Plasmodium knowlesi* in Malaya. *Science*, 134(3474), 279 –280.
- Wharton, R. H., Eyles, D. E., and Moorhouse, D. E. (1962). *Anopheles leucosphyrus* identified as a vector of monkey malaria in Malaya. *Science*, 137(3532), 758. doi: 10.1126/science.137.3532.758
- Wharton, R. H., Eyles, D. E., Warren, Mc.W., and Cheong, W. H. (1964). Studies to determine the vectors of monkey malaria in Malaya. *Annals of Tropical Medicine and Parasitology* 58, 56-77. World Health Organisation (2008).
- White, G. B. (1982). Malaria vector ecology and genetics. *British Medical Bulletin*, 38(2), 207–212.
- White, N. J. (2008). *Plasmodium knowlesi*: the fifth human malaria parasite. *Clinical Infectious Diseases*, 46(2), 172-173. doi: 10.1086/524889

- White, N. J. (2011). Determinants of relapse periodicity in *Plasmodium vivax* malaria. *Malaria Journal*, 10(1), 297. doi:10.1186/1475-2875-10-297
- White, N. J., Pukrittayakamee, S., Hien, T. T., Faiz, M. A., Mokuolu, O. A., and Dondorp, A. M. (2014). Malaria. *Lancet*. 383, 723-735. doi:10.1016/S0140-6736(13)60024-0
- Wilfinger, W. W., Mackey, K., and Chomczynski, P. (1997). Effect of pH and ionic strength on the spectrophotometric assessment of nucleic acid purity. *BioTechniques*, 22(3), 474-6.
- William, T., Menon, J., Rajahram, G., Chan, L., Ma, G., Donaldson, S., Khoo, S., Frederick, C., Jelip, J., Anstey, N. M., and Yeo, T. W. (2011). Severe *Plasmodium knowlesi* malaria in a tertiary care hospital, Sabah, Malaysia. *Emerging Infectious Diseases*, 17(7), 1248-55. doi: 10.3201/eid1707.101017
- William, T., Rahman, H. A., Jelip J, Ibrahim, M. Y., Menon, J., Grigg, M. J., Yeo, T. W., Anstey, N. M, and Barber, B. E. (2013). Increasing incidence of *Plasmodium knowlesi* malaria following control of *P. falciparum* and *P. vivax* Malaria in Sabah, Malaysia. *PLoS Neglected Tropical Diseases*, 7(1), e2026.
- William, T., Jelip, J., Menon, J., Anderios, F., Mohammad, R., Mohammad, T. A. A., Grigg, M. J., Yeo, T. W., Anstey, N. M. and Barber, B. E., (2014). Changing epidemiology of malaria in Sabah, Malaysia: increasing incidence of *Plasmodium knowlesi*. *Malaria Journal*, 13, 390. doi:10.1186/1475-2875-13-390
- Winzeler, E. A. (2008). Malaria research in the post-genomic era. *Nature*, 455(7214), 751-756. doi:10.1038/nature07361
- Wongsrichanalai, C., Barcus, M. J., Muth, S., Sutamihardja, A., and Wernsdorfer, W. H. (2007). A review of malaria diagnostic tools: Microscopy and rapid diagnostic test (RDT). *American Journal of Tropical Medicine and Hygiene*, 77(6), 119–127.
- World Health Organization. (2012). *Defeating Malaria in Asia, the Pacific, Americas, Middle East and Europe*. World Health Organization.
- World Health Organization. (2013). World Malaria Report 2013. Geneva, Switzerland.  
Retrieved from:  
[http://www.who.int/malaria/publications/world\\_malaria\\_report\\_2013/report/en/](http://www.who.int/malaria/publications/world_malaria_report_2013/report/en/).
- World Health Organization. (2014). World Malaria Report 2014. Geneva, Switzerland.  
Retrieved from:  
[http://www.who.int/malaria/publications/world\\_malaria\\_report/en/](http://www.who.int/malaria/publications/world_malaria_report/en/)
- World Health Organization (2014). List of known commercially available antigen-detecting malaria RDTs. Retrieved from: <http://www.wpro.who.int/sites/rdt>

- Wright, S. (1978). Evolution and the genetics of populations. Variability within and among natural populations. Vol. 4. *University of Chicago press, Chicago, IL, USA.*
- Yakob, L., Bonsall, M. B., and Yan, G. (2010). Modelling *knowlesi* malaria transmission in humans: vector preference and host competence. *Malaria Journal*, 9(1), 329. doi:10.1186/1475-2875-9-329
- Ying, P., Shakibaei, M., Patankar, M. S., Clavijo, P., Beavis, R. C., Clark, G. F., and Frevert, U. (1997). The malaria circumsporozoite protein: interaction of the conserved regions I and II-plus with heparin-like oligosaccharides in heparan sulfate. *Experimental Parasitology*, 85(2), 168-182. doi:10.1006/expr.1996.4134
- Yusof, R., Lau, Y. L., Mahmud, R., Fong, M. Y., Jelip, J., Ngian, H. U., and Ali, M. M. (2014). High proportion of *knowlesi* malaria in recent malaria cases in Malaysia. *Malaria Journal*, 13(1), 168. doi:10.1186/1475-2875-13-168.
- Zeeshan, M., Alam, M. T., Vinayak, S., Bora, H., Tyagi, R. K., Alam, M. S., Choudhary, V., Mittra, P., Lumb, V., Bharti, P. K. and Udhayakumar, V., (2012). Genetic variation in the *Plasmodium falciparum* circumsporozoite protein in India and its relevance to RTS, S malaria vaccine. *PloS one*, 7(8), e43430. doi:10.1371/journal.pone.0043430
- Zhu, H. M., Li, J., and Zheng, H. (2006). Human natural infection of *Plasmodium knowlesi*. *Zhongguo Ji Sheng Chong Xue Yu Ji Sheng Chong Bing Za Zhi* 24(1), 70 –71.
- Zilversmit, M., Hartl, D. L., and Sherman, I. W. (2005). Evolutionary history and population genetics of human malaria parasites. *Molecular Approaches to Malaria*, 95-109.